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**EFFECT OF ENHANCED LEVELS OF CARBON DIOXIDE  
ON PRIMARY PRODUCTIVITY**

Dissertation submitted to the Jawaharlal Nehru University  
in partial fulfilment of the requirements  
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**1991**

*To  
My family*



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CERTIFICATE

The research work embodied in this dissertation entitled "Effect of enhanced levels of carbon dioxide on primary productivity" has been carried out in the School of Environmental Sciences, Jawaharlal Nehru University, New Delhi. This work is original and has not been submitted so far, in part or full, for any other degree or diploma of any University.

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## PART I: EFFECT OF ENHANCED LEVEL OF CO<sub>2</sub> ON PLANTS

Carbon dioxide is one of the natural constituents, comprising 0.03% of the earth's atmosphere. Plants take up CO<sub>2</sub> and assimilate carbon by the process of photosynthesis creating reduced carbon (C<sub>n</sub>H<sub>2n</sub>O). The reduced carbon is stored in biota, in dead organic matter in soil and in the top layers of the sediments, in coal, oil and gas reserves, and as highly dispersed carbon in the lithosphere. Expanding human activities involving fossil fuel combustion and large scale deforestation have resulted in increased levels of CO<sub>2</sub> in the atmosphere.

The atmospheric concentration of CO<sub>2</sub> is now 25% higher than during the first half of the last century. (Houghton and Woodwell, 1989). Direct sampling of air at the South Pole and Mauna Loa, Hawaii, has shown that the CO<sub>2</sub> concentration of the atmosphere has risen from about 314 ppm in 1958 to over 345 ppm today (Keeling et al., 1989). This change is due to, the burning of fossil-fuels which is ejecting approximately 5-6 g T (1 giga tonne = 10<sup>9</sup> tonne) of carbon into the atmosphere yearly, and deforestation which may account for another 1-2gT (Moore and Bolin, 1986/87 ; Detwiler and Hall, 1988). Future projections indicate that by the year 2065, atmospheric CO<sub>2</sub> levels will reach 600 ppm (IPCC, 1990).

In addition to carbon dioxide other atmospheric trace gases such as methane, ozone, nitrous oxide and chlorofluorocarbons (CFC<sub>11</sub> and CFC<sub>12</sub>) are transparent to incoming solar radiation but absorb the out going infra-red radiation from the earth. This phenomenon is known as the 'greenhouse effect' and the gases causing it are known as the greenhouse gases (Jager, 1986). Increasing levels of greenhouse gases are expected to bring serious changes in global climate (Jager, 1986). The focus of this review is to evaluate the effect of enhanced levels of CO<sub>2</sub>, on plants at individual and ecosystem levels.

Rising levels of CO<sub>2</sub> can affect plants in two ways, (i) directly due to higher concentration of ambient CO<sub>2</sub> on plant growth and development or (ii) indirectly due to global climate change triggered by rising levels of CO<sub>2</sub> manifested as rise in average ambient temperature, alteration in precipitation regimes and associated changes (Warrick et al., 1984). In general, higher ambient CO<sub>2</sub> stimulates greater net photosynthesis, the so called CO<sub>2</sub> fertilization effect. High CO<sub>2</sub> levels suppress transpiration through partial closure of stomata, resulting in greater water use efficiency (Warrick et al., 1984). However, the extrapolation from individual plants to dynamic ecosystems is highly tenuous. Competition between plants, and

consequent interaction between plants, animals and microbes are likely to change drastically (Warrick et al., 1984).

A cursory, global survey of natural systems reveals an unmistakable correspondence between the broad features of regional climates and the major characteristics of world's biomes. Major changes in the global climate will bring about major changes in natural biomes which will be particularly more dramatic in transtion zones or ecotones (Warrick et al., 1984).

#### **A. SPECIES LEVEL RESPONSE:**

Species level responses to elevated CO<sub>2</sub> have been discussed in relation to herbaceous, tree and aquatic plant species.

##### **A.1 HERBACEOUS SPECIES**

Predictions of crop growth and yield under elevated CO<sub>2</sub> are incomplete if based solely on photosynthetic response at the level of primary CO<sub>2</sub> fixation mechanism. Other primary and secondary responses (like stomatal conductance and morphological development) and feedbacks interpose between photosynthetic metabolism and crop yield and must be taken into consideration in assessing the effects of higher atmospheric CO<sub>2</sub> in particular a CO<sub>2</sub> doubling (Warrick et al., 1984).

### Growth and Development:

**Leaf area :** In several C<sub>3</sub> species, leaf area has been found to increase under elevated CO<sub>2</sub> conditions in response to improved photosynthate supply (Rogers et al., 1984; Delucia et al., 1985). Soybean (Glycine max) grown at twice normal CO<sub>2</sub> showed an increased leaf area (Rogers et al., 1984). Elevated CO<sub>2</sub> had a small accelerating effect on the rate of leaf initiation and also caused a faster expansion of the G. max leaves (Rogers et al., 1986). In cotton, (Gossypium hirsutum) CO<sub>2</sub> enrichment caused a significant increase in leaf area (Delucia et al, 1985) in the first 20 days, following emergence, (Mauney et al., 1978). CO<sub>2</sub> enrichment resulted in increased total canopy area, 36% greater than controls on day 22, in soybean (Cure et al., 1987). In sugarbeet (Beta vulgaris) (Wyse, 1980) increased CO<sub>2</sub> also increased leaf number in addition to leaf area increase. In bean (Phaseolus vulgaris) plants (Porter and Grodzinski, 1984), there was an increase in leaf area only after 14 days of CO<sub>2</sub> enrichment. On 14th day in ambient treatment leaf area was 144.3 cm<sup>2</sup>/plant compared to 183.6 cm<sup>2</sup>/plant under high CO<sub>2</sub>. Rice (Oryza sativa) does not increase leaf area appreciably under CO<sub>2</sub> enrichment even though dry weight growth responds (Yoshida, 1972; Morison and Gifford, 1984).

Conversely, several C<sub>4</sub> species that did not show a response of net CO<sub>2</sub> fixation per unit leaf area or per unit of intercepted radiation, nevertheless responded with an increase in leaf area (Patterson and Flint, 1980; Morison and Gifford, 1984b). Growth analysis of both maize (Zea mays) and itchgrass (Rottboellia exaltata) (Patterson and Flint, 1980) showed that leaf area increased while NAR (net assimilation rate) was unaffected by CO<sub>2</sub> enrichment to above 600 ppm. Similarly with a doubling of normal CO<sub>2</sub>, Morison and Gifford (1984b) observed increases in leaf area of the C<sub>4</sub> species Amaranthus edulis (15%), Sorghum bicolor (29%), and Zea mays (40%), NAR remaining unchanged by high CO<sub>2</sub>. Thus, the increase in growth by higher CO<sub>2</sub> in these C<sub>4</sub> species was attributable to greater interception of light because of bigger leaf area, not to increased photosynthesis per unit leaf area, implying that CO<sub>2</sub> was acting on leaf area development in some way other than via CO<sub>2</sub> effects on photosynthesis rate (Morison and Gifford, 1984).

The mechanisms involved in CO<sub>2</sub> - stimulated leaf area expansion have not been widely investigated. Depending on the species, the component of leaf area increase varies between axillary growth, faster rate of leaf emergence and development of larger leaves (Goudriaan and de Ruiter, 1983).

**Growth rate** : Increased growth rates at high CO<sub>2</sub> levels may be caused by a number of factors (Mott, 1990). These could be, an increased supply of photosynthetically fixed carbon, increased plant water potential and turgor due to stomatal closure, increased osmoregulation under water stress due to improved carbon supply, decreased dark respiration rates or direct effect of CO<sub>2</sub> on rates of cell division or enlargement (Mott, 1990).

Growth rates are generally enhanced when elevated CO<sub>2</sub> is given in the early stages of growth (Thomas et al., 1975; Mauney et al., 1978). But with small grain cereals like wheat (Triticum aestivum), interesting results have been obtained. An increase in grain yield (36%) with a doubling of CO<sub>2</sub> is nearly twice the increase in biomass of immature crops (20%) (Kimball, 1983). The effects of high CO<sub>2</sub> on wheat seedlings is small (Neales and Nicholls, 1978) compared to the effects once tillering and grain formation occur (Gifford, 1977; Sionit et al., 1981a).

Despite decreases in photosynthetic rates and relative growth rates during long-term CO<sub>2</sub> enrichment, plants grown at high CO<sub>2</sub> concentration usually continue to grow at a faster absolute rate and maintain higher dry weight over control plants throughout the enrichment period. This has been attributed to the fact that increases

in growth rate during the early period of growth increase the leaf area over that of controls. This allows higher rates of photosynthesis per plant and a higher growth rate, despite similar rates of photosynthesis per leaf area (Spencer & Bowes, 1986; Curtis et al., 1989a; Mauney et al., 1978).

**Biomass :** Cure and Acock (1986) calculated that the average increase in biomass for C<sub>3</sub> grasses grown under twice normal CO<sub>2</sub> concentration was 28% but increases of 100% or more in biomass have been reported for some C<sub>3</sub> plants (Delucia et al., 1985; Mauney et al., 1978; Wong, 1979). Sage et al. (1989) found that in five C<sub>3</sub> species, they studied, growth at high CO<sub>2</sub> significantly increased leaf dry weight per area, probably due to accumulation of starch. Clough et al. (1981), experimenting with soybean plants, observed that under high CO<sub>2</sub> vegetative dry weights and per pod dry weights were higher compared to those under ambient CO<sub>2</sub>. Bean (*P. vulgaris*) plants exposed to high CO<sub>2</sub>, (Porter and Grodzinski, 1984) after 14 days, showed a 71% increase in dry weight, and specific leaf weight was also found to increase. Fresh and dry weights of whole cowpea (*Vigna unguiculata*) plants increased at elevated CO<sub>2</sub> (640 ppm) concentrations (Mbikayi et al., 1988) as compared to plants grown at ambient CO<sub>2</sub>. Patterson and

Flint (1980) studied dry matter production in two C<sub>3</sub> species (G. max and Abutilon theophrasti) and two C<sub>4</sub> species (Z. mays and R. exaltata). C<sub>3</sub> dry weight growth responded to CO<sub>2</sub> concentrations above 350 ppm whereas C<sub>4</sub> species did not respond. In Pisum sativum (Paez et al., 1980) not much difference in total plant dry weights was observed at high CO<sub>2</sub> (1000 ppm) and ambient CO<sub>2</sub> (350 ppm) even after 39 days of exposure. In B. vulgaris, Wyse (1980) observed a 180% increase in total dry weight over a 10 day exposure to 1000 ppm CO<sub>2</sub> compared to ambient. Delucia et al. (1985) found a similar increase in dry weight in G. hirsutum plants grown at 675 ppm (72% increase) and 1000 ppm CO<sub>2</sub> (115% increase) over that of plants grown at 350 ppm CO<sub>2</sub>. Cure et al. (1987) observed a 69% increase in dry weight of high CO<sub>2</sub> (700 ppm) grown G. max plants over a 3-week period. It seems probable that the increase in dry weight in C<sub>3</sub> plants under elevated CO<sub>2</sub> is largely the result of increased photosynthetic assimilation whereas C<sub>4</sub> plants do not respond to elevated CO<sub>2</sub> in this respect.

**Flowering :** No particular trend in the onset of flowering in relation to elevated CO<sub>2</sub> concentration is discernible. Amaranthus retroflexus flowered significantly earlier at 700 ppm CO<sub>2</sub> than at 350 ppm, whereas Setaria faberii flowered significantly later at 700 ppm CO<sub>2</sub> (Garbutt et al., 1990). A. theophrasti and Ambrosia



artemisiifolia showed a trend towards earlier flowering at high CO<sub>2</sub> (Garbutt et al., 1990). A slowing in the rate of flower development in Sorghum under elevated CO<sub>2</sub> without any change in dry weight growth (Marc & Gifford, 1983) has been observed. There is no conclusive evidence that dry weight is preferentially allocated to reproductive structures as fruits and flowers. Calculated yield increases for CO<sub>2</sub> enrichment of agricultural species (Kimball, 1983; Cure & Acock, 1986) are not different from increases in total biomass with CO<sub>2</sub> enrichment indicating that carbon is allocated more or less equally among reproductive and vegetative portions of the plant. However investigations in this area would be most interesting. Study of effects of CO<sub>2</sub> enrichment on morphology and functioning of floral parts would provide us with a better insight into the mechanism(s) underlying the observed responses.

**Senescence** : An increased rate of senescence (aging) due to CO<sub>2</sub> enrichment, has been widely reported in the literature. Accelerated senescence has been observed in G. hirsutum (Chang, 1975), under 850 and 1000 ppm CO<sub>2</sub>. But Carter and Peterson (1983) observed delayed senescence in Sorghum at 600 ppm CO<sub>2</sub>. Curtis et al. (1989a) observed a decreased rate of senescence in the C<sub>3</sub> sedge Scirpus olneyi. Although the observed senescence effect is minor, and is not

always detected (eg. no effect in wheat; Gifford, 1977), it could possibly be pervasive due to increase in ethylene, a natural growth regulator in plants which accelerates senescence. High CO<sub>2</sub> concentrations caused H. annuus shoots to produce more ethylene, (Dhawan et al., 1981). In addition, the CO<sub>2</sub> source for enriching the air might also contain unsuspected traces of ethylene which could promote early senescence (Morison and Gifford, 1984a). Early senescence under elevated CO<sub>2</sub> may also be correlated with the timing of other phenological events such as flowering (St. Omer and Horvath, 1983).

#### Physiology and Biochemistry

**Stomatal conductance** : The presence of a CO<sub>2</sub> - impermeable cuticle on most aerial surfaces of land plants makes the direct sensing of atmospheric CO<sub>2</sub> unlikely. Most of the known responses to atmospheric CO<sub>2</sub> can be attributed to changes in intercellular CO<sub>2</sub> (C<sub>i</sub>) concentration (Mott 1988) and more specifically to the effect of changes in C<sub>i</sub> on stomatal conductance (Mott, 1990). Changes in ambient CO<sub>2</sub> concentration will cause changes in C<sub>i</sub>, such that the ratio of ambient CO<sub>2</sub> to C<sub>i</sub> remains approximately constant (Bell, 1982). The general trend in response of stomatal conductance to increasing CO<sub>2</sub> concentrations is that of a decreasing one (Cure and Acock, 1986). In soybean (Rogers et

al., 1984), stomatal conductance measurements have been shown to decrease significantly, from  $1.0 \text{ cm s}^{-1}$  (300 ppm) to  $0.25 \text{ cm s}^{-1}$  (900 ppm). In cotton (Delucia et al., 1985), a short term exposure to  $\text{CO}_2$  caused stomatal conductance to decline from  $0.6 \text{ cm s}^{-1}$  (350 ppm) to  $0.4 \text{ cm s}^{-1}$  (100 ppm). In V. unguiculata (Mbikayi et al., 1988), stomatal conductance on adaxial surface was lower than abaxial surface in leaves, but decreased in both cases at 655 ppm  $\text{CO}_2$  compared to 354 ppm. During long-term exposures to elevated  $\text{CO}_2$ , a further decline in stomatal conductance values has been observed (Spencer & Bowes, 1986). There is no difference between  $\text{C}_3$  and  $\text{C}_4$  plants with respect to the sensitivity of stomatal conductance to change in  $\text{CO}_2$  concentration (Morison and Gifford, 1983). A reasonable approximation is that, for most species and environmental conditions, a  $\text{CO}_2$  doubling will cause about a 34% decrease in stomatal conductance (Cure and Acock, 1986).

The mechanism for stomatal responses to  $\text{CO}_2$  is unknown at present (Mott, 1990). The existence and functioning of the photosynthetic carbon reduction cycle in guard cell chloroplasts is controversial (Tarczynski et al., 1989), however it is possible that stomatal sensitivity to  $\text{CO}_2$  may depend on the response of photosynthesis to  $\text{CO}_2$  in stomatal guard cells (Mott, 1990). Stomatal responses to  $\text{C}_i$  have evolved to compensate for changes in  $\text{C}_i$  caused by

changes in mesophyll demand for CO<sub>2</sub>, they may not regulate gas exchange optimally for changes in C<sub>i</sub> caused by an increase in ambient CO<sub>2</sub> (Mott, 1990). Other aspects of stomatal physiology including the effects of CO<sub>2</sub> enrichment on stomatal responses to light and humidity, are areas which have to be probed in order to define an optimal response of stomata to enriched ambient CO<sub>2</sub> concentrations (Mott, 1990).

**Chlorophyll content :** A generally decreasing trend in chlorophyll content with increasing CO<sub>2</sub> concentration has been observed for most of the species studied. Madsen (1968) reported, no variation in chlorophyll content in tomato (Lycopersicum esculentum) when measured on a leaf area basis, but on a fresh weight basis reduction in chlorophyll was observed at elevated level of CO<sub>2</sub>. In G. hirsutum (Chang, 1975), about 42% decrease in chlorophyll content on fresh weight basis was observed at 1000 ppm CO<sub>2</sub> compared to ambient CO<sub>2</sub>. In Trifolium subterraneum (Cave et al., 1981), total chlorophyll, calculated on a dry weight basis, in immature leaves was 34% lower in 1000 ppm treatment compared to 350 ppm. Mature leaves showed a 30% decrease in chlorophyll. Chlorophyll a : b ratio also decreased for high CO<sub>2</sub> plants, 22% decrease in immature leaves and a 33% decrease in mature leaves (Cave et al., 1981). Delucia et al. (1985) also reported a 61% decrease in total

chlorophyll on dry weight basis and a decreased Chl a:b ratio in cotton at elevated CO<sub>2</sub> (1000 ppm) compared to 350 ppm grown plants. Sage et al. (1989) reported a decline in chlorophyll content at 950 ppm CO<sub>2</sub> in Chenopodium album (14%) and Brassica oleracea (34%) compared to ambient.

The decrease in chlorophyll content associated with increasing atmospheric CO<sub>2</sub> could be due to chloroplast degeneration as a result of excess starch accumulation (Madsen, 1968). The electron micrographs offer additional evidence that increasing starch accumulation in plants growing in enriched CO<sub>2</sub> atmospheres affects chloroplast structure and whole plant chlorophyll content, contributing to chlorosis of leaves. (Cave et al., 1981). The lower chlorophyll a:b ratio is primarily accounted for by a reduction in chlorophyll (a) content and not an increase in chlorophyll (b) (Cave et al., 1981). Decline in chlorophyll content could thus place serious limitations on the photosynthetic capacity of plants in the long run.

**Photosynthesis** : Photosynthesis plays a central role in the physiology of plants. Thus, it is likely that many responses exhibited by plants to elevated CO<sub>2</sub> are, in fact, mediated by response of photosynthesis to elevated CO<sub>2</sub>. C<sub>3</sub> and C<sub>4</sub> plant photosynthesis has been reported to respond to elevated CO<sub>2</sub> in a strikingly different manner (Pearcy and Ehleringer, 1984).

Short term exposure of C<sub>3</sub> plants to elevated CO<sub>2</sub> typically causes an increase in the rate of net photosynthesis. In contrast several studies have shown that long term exposure can result in a subsequent decline in net carbon assimilation when measured on a leaf area basis. Cure and Acock (1986) calculated that photosynthesis is initially stimulated at an average of 52% after doubling the CO<sub>2</sub> concentration. But this average increase is only 29% after plants acclimate to new CO<sub>2</sub> concentration. Tobacco (Nicotiana tabacum) plants grown at 1000 ppm CO<sub>2</sub> for a period of 35 days showed a 20% decline in the rate of net photosynthesis. (Raper and Peedin, 1978). In another study, long term exposure (4 weeks) of cotton plants to 1000 ppm CO<sub>2</sub> caused a decline in net photosynthesis after an initial increase. The 350 ppm plants consistently had higher rates of photosynthesis than the 675 ppm or 1000 ppm CO<sub>2</sub> plants. (Delucia et al., 1985). In the C<sub>3</sub> species G. max and A. theophrasti (Patterson and Flint, 1980) increasing CO<sub>2</sub> concentration from 350 to 600 ppm increased the NAR (Net assimilation rate) by 35%. In C<sub>4</sub> species, Z. mays and R. exaltata, elevated CO<sub>2</sub> did not affect the NAR, (Patterson and Flint, 1980). In a C<sub>3</sub> sedge, Scirpus olneyi (Ziska et al, 1990), however, increased photosynthetic rates were maintained throughout the two years of experiment, without an acclimation to high CO<sub>2</sub>.

The observed increase in net photosynthesis in  $C_3$  plants under  $CO_2$  enrichment could be due to an improved competitive advantage of  $CO_2$  molecules over  $O_2$  molecules for the active sites on rubisco. The reduced carbon flow through the photorespiratory cycle leads to less photorespiratory  $CO_2$  loss as well. Hence  $C_3$  plants are expected to respond positively to elevated  $CO_2$  atmospheres (Warrick et al., 1984). In contrast, the primary carboxylase in  $C_4$  plants is PEP carboxylase which is not competitively inhibited by  $O_2$ . Photorespiration is therefore negligible. PEP carboxylase has a higher effective affinity for  $CO_2$  than does rubisco in the absence of  $O_2$ , so the enzyme is close to  $CO_2$  saturation at the present atmospheric  $CO_2$  concentration. Also in  $C_4$  plants rubisco is located in the bundle sheath cells, where the  $CO_2$  concentration largely saturates carboxylation and inhibits oxygenation. Therefore, one would not expect a significant enhancement of  $C_4$  crop growth from increased  $CO_2$  in so far as the primary carboxylase properties are concerned (Warrick et al., 1984).

The mechanism (s) responsible for the decrease in photosynthetic rate over long term exposure have not been established clearly. Under long term exposures, the activity of growth sinks and the associated ability to utilise the increased supply of photosynthate plays an important role (Clough et al., 1981). If sink demand is insufficient,

assimilates can accumulate in source leaves, resulting in end product inhibition of photosynthesis. Soybean plants in which pod set had taken place were taken and trimmed to either 21 pods (high sink) or 6 pods (low sink) (Clough et al., 1981). Comparing plants from the same CO<sub>2</sub> treatment, high sink plants had greater rates of photosynthesis at 1000 ppm CO<sub>2</sub> than the low sink plants. Thus, high source : sink ratios are associated with lower rates of photosynthesis. The more rapidly storage tissues are filled the more rapidly rates of photosynthesis decline. Mechanism for feedback inhibition of photosynthesis is supported by measurements showing increased levels of starch and sucrose in many plants subjected to prolonged CO<sub>2</sub> enrichment (Spencer and Bowes, 1986). The degree of starch accumulation in plants grown in high CO<sub>2</sub> is often so great that distortion of chloroplasts by starch grains has been suggested as a mechanism for decreasing the rate of photosynthesis (Madsen, 1968, Cave et al., 1981).

A decline in rubisco activity under elevated CO<sub>2</sub> could be another factor in acclimation of plants to high CO<sub>2</sub>. At normal ambient CO<sub>2</sub> concentration, photosynthesis is limited significantly by RUBP regeneration capacity, and sucrose synthesis capacity (Von Caemmerer and Farquhar, 1984). When ambient CO<sub>2</sub> is increased, however, the



apparent maximum catalytic capacity of rubisco is also increased and the balance among limitations is upset. RUBP regeneration and sucrose synthesis become more limiting and carboxylation capacity (Rubisco) becomes less limiting (Von Caemmerer and Farquhar, 1984). In view of the altered balance among these three limiting elements, it has been hypothesized (Sage et al, 1989) that acclimation of the photosynthetic system to high  $\text{CO}_2$  should involve re-allocation of protein nitrogen from rubisco to the enzymes of light harvesting, RUBP regeneration and starch and sucrose synthesis. The effects of such a re-allocation would be to restore the balance among limiting factors. Since rubisco constitutes the single largest sink for N in the photosynthetic apparatus, changes in its content will have greatest effect on N partitioning within the leaf. Rubisco activity does decline following long term exposure to high  $\text{CO}_2$ . In B. oleracea and in C. album, (Sage et al., 1989) leaf rubisco content was lower in plants grown at high  $\text{CO}_2$  (950 ppm). The percent of leaf N invested in rubisco was lower in plants grown at high  $\text{CO}_2$ , particularly C. album. The rubisco activation state was also lower in leaves of all five species grown at high  $\text{CO}_2$  (Sage et al, 1989). In P. vulgaris too, rubisco activity dropped by 40% under elevated  $\text{CO}_2$ . Thus, a decline in rubisco activity under elevated  $\text{CO}_2$  plays an important role in acclimation of plants under high

CO<sub>2</sub>. There is no evidence, however, to suggest that plants re-allocate nitrogen to relieve limitations by starch and sucrose synthesis. Sucrose synthase and sucrose - P - synthase activities in plants grown at high CO<sub>2</sub> were found to be similar to those in plants grown at normal CO<sub>2</sub> (Peet et al, 1986).

**Photosynthate partitioning:** Ambient CO<sub>2</sub>, level seems to play a definite role in photosynthate partitioning (Wyse, 1980). Reports on biomass partitioning under elevated CO<sub>2</sub> have shown mixed results.

In soybean (Cure et al, 1987) by day 22 of exposure to 700 ppm CO<sub>2</sub>, dry weight of leaves increased 60%, stems 73% and roots 88% above the controls indicating preferential allocation to roots. In B. vulgaris (Wyse, 1980), the additional photosynthate resulting from enhanced photosynthesis at elevated CO<sub>2</sub> (1000 ppm), was allocated preferentially to root sink. In Bromus mollis root: shoot ratio increased at elevated CO<sub>2</sub> mainly due to increase in root biomass (Larigauderie et al., 1990). The partitioning of biomass between roots and shoots was not affected by CO<sub>2</sub> concentration in the C<sub>4</sub> species, Z. mays and R. exaltata (Patterson and Flint, 1980) However, in the C<sub>3</sub> species, (G. max., A. theophrasti) the root : shoot ratios tended to increase with increasing CO<sub>2</sub> concentration (Patterson and

Flint, 1980). These effects of CO<sub>2</sub> enrichment on dry matter partitioning between roots and shoots may have implications for weed - crop competitive interactions. C<sub>3</sub> weeds will become more competitive with crops having C<sub>4</sub> pathway. Weeds with C<sub>4</sub> pathway may become less competitive with crops having C<sub>3</sub> pathway (Patterson and Flint, 1980). In another study on soybean (Finn and Brun, 1982), it was observed that additional photosynthate provided by CO<sub>2</sub> enrichment was being utilized predominantly by shoot material for growth and storage, with relatively little being partitioned to the roots and nodules. This result is contrary to observations on soybean noted above. In cotton (Delucia et al., 1985) too, biomass partitioning was preferential to the leaf sink followed by stems and least to the roots.

Biomass partitioning thus seems to be guided by individual plant characteristics and requirements and does not seem to follow a general pattern under CO<sub>2</sub> enrichment.

**Starch content** : Elevated CO<sub>2</sub> treatment has a profound effect on the diurnal pattern of leaf starch accumulation (Delucia et al., 1985). Under ambient CO<sub>2</sub> levels, starch concentration gradually increases throughout the light period and declines to the previous morning's levels by the end of the dark period (Delucia et al., 1985). In elevated CO<sub>2</sub> (675 and 1000 ppm) grown cotton plants

(Delucia et al., 1985), the rate of increase and maximum starch concentration during the light period was considerably greater. Due to insufficient translocation and or degradation of carbohydrates in high CO<sub>2</sub> grown plants, the starch pool did not return to the previous morning's level by the end of dark period. A similar behaviour was observed in tomato plants by Madsen (1968). At elevated CO<sub>2</sub> concentrations starch content maxima is reached within 1-2 hr after sunrise, whereas for control plants, it is reached only at noon. Cave et al. (1981) observed that, in T. subterraneum leaves, there was a significant increase in starch content in late afternoon as compared to early morning. In high CO<sub>2</sub> plants the increase was 135% whereas in control plants it was 46.7% only. In soybean (Finn and Brun, 1982). majority of the additional carbohydrates provided by CO<sub>2</sub> enrichment were stored in the shoots as leaf starch, resulting in 46% increase in foliar starch content.

In most C<sub>3</sub> plants, CO<sub>2</sub> enrichment produces a large increase in starch concentrations, causing a disruption of equilibrium in the starch pool size on a diurnal basis. This disequilibrium in pool size would grow with each day, unless and until degradation and/or translocation to sinks, of this additional starch takes place. CO<sub>2</sub> enrichment has been found to have little or no effect on concentration of soluble sugars resulting from

degradation of additional starch (Madsen, 1968). Sink size and number has been found to increase under CO<sub>2</sub> enrichment for few plants like wheat, rice, soybean (Gifford, 1979; Cock and Yoshida, 1973; Finn and Brun, 1982).

Starch accumulation can limit the rate of photosynthesis by feedback inhibition during long term exposure, thus putting constraints on the increase in productivity under high CO<sub>2</sub>.

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Respiration : Experiments on the effects of high CO<sub>2</sub> concentrations on respiration show mixed results. It has been proposed that mitochondrial respiration may increase in plants under high CO<sub>2</sub> in response to sucrose accumulation in leaves (Tolbert et al., 1983). A mechanism for this is thought to act via the alternative pathway of respiration, that may function to dissipate excess photosynthesized energy (Lambers, 1982). Hrubec et al. (1984), reported increased respiration rates of soybean leaves grown in high CO<sub>2</sub>. However, the converse result was found for wheat (Gifford et al., 1985) plants grown in 590 ppm CO<sub>2</sub>, which experienced up to 45% reduction in respiration by both roots and whole plants. The operation of the alternative pathways of respiration actually declined for continuously CO<sub>2</sub> enriched plants. For V. radiata (Gifford et al., 1985) high CO<sub>2</sub> caused no significant change in root respiration per



unit root dry weight. H. annuus (Gifford et al., 1985) had another pattern of response. Here root respiration per unit dry weight was increased, this increase was not attributable to the alternative pathway but to the cytochrome oxidase pathway. Increasing CO<sub>2</sub> from 350 to 950 ppm for a short term reduced the rate of dark respiration of Medicago sativum (Reuveni and Gale, 1985), the suppression being greater for roots than tops. The above short term effect of high CO<sub>2</sub> was confirmed for longer periods. High CO<sub>2</sub> at night (~ 1000 ppm) reduced respiration and increased the 24 hr net carbon gain and calculated dry weight growth. In these, as for short term experiments, the percentge reduction of respiration was greater when the plant was in a low photosynthate "maintenance respiration" condition (28% vs 11%) (Reuveni and Gale, 1985).

The mechanism of action of high CO<sub>2</sub> on dark respiration is not clear. Either high CO<sub>2</sub> directly affects respiration or CO<sub>2</sub> might affect rate of an energy requiring process whose ATP or NADPH demand affects respiration. These results suggest that even a small depression of respiration may have a significant effect on growth if two conditions are met (Reuveni and Gale, 1985):

- (1) that the plant is growing under conditions in which the net daily carbon gain is low and
- (2) reduction of respiration is not deleterious.

Thus this effect will be especially pronounced in regions and under conditions in which the daily net carbon gain is low or negative, for example in areas receiving low insolation at high latitudes or low water availability in arid regions (Reuveni and Gale, 1985).

**Carbon and nitrogen ratio :** Elevated CO<sub>2</sub> levels are expected to have an impact on the elemental composition of plant tissues, particularly the carbon and nitrogen content. C/N ratios under high CO<sub>2</sub> are likely to increase.

Curtis et al. (1989b) exposed three plant communities on a brackish marsh of Rhode river, to elevated CO<sub>2</sub> concentrations for an entire growing season. Two communities were monospecific one of S. olneyi (C<sub>3</sub>) other of Spartina patens (C<sub>4</sub>), the third was a mixed community of S. olneyi, S. patens and Distichlis spicata(C<sub>4</sub>). A clear dichotomy was observed in the effects of elevated CO<sub>2</sub> on shoot % N in the C<sub>3</sub> and C<sub>4</sub> species. Elevated CO<sub>2</sub> reduced green tissue % N in the C<sub>3</sub> sedge S.olneyi but had no effect on the C<sub>4</sub> grasses S. patens or D. spicata. Percent carbon varied only slightly. This decrease in tissue % N in Scirpus caused a significant increase in C/N ratio of about 20-40% in pure as well as mixed community. There was no difference in % C or % N of seeds from Scirpus between elevated and ambient treatments. The enveloping bracts,

however, had significantly less % N under elevated CO<sub>2</sub>. Total litter N, while unaffected by CO<sub>2</sub> in Scirpus in pure stand, increased significantly in Scirpus from mixed community (Curtis et al., 1989b).

Chang (1975) while working on G. hirsutum found that under elevated CO<sub>2</sub>, protein content declined in the leaves with no accompanying increase in ninhydrin positive compounds. This observation evidenced that high CO<sub>2</sub> decreases the content of protein, not by degradation, but by curtailing protein synthesis. Mbikayi et al. (1988), found that in V. unguiculata, after 41 days of exposure in elevated CO<sub>2</sub> there was no effect on protein content of either shoots or roots. After 79 days of exposure protein nitrogen content of both shoot and seeds was not affected by increasing levels of CO<sub>2</sub>, but that of roots decreased significantly.

Sage et al. (1989) studied five C<sub>3</sub> species, (C. album, P. vulgaris, Solanum tuberosum, Solanum melongena and B. oleracea) for various parameters under elevated CO<sub>2</sub>. With respect to leaf nitrogen content per unit area, they found that it increased in two Solanum species but was little changed in the other three. Nitrogen per unit weight fell in all species following exposure to high CO<sub>2</sub>, but this was largely a consequence of the increase in leaf weight per



area. Leaf N content was found to decline under elevated CO<sub>2</sub> (500 and 700ppm) in all five annuals studied: A. theophrasti, A. retroflexus, A. artemisiifolia, C. album and S. faberii (Garbutt et al., 1990). B. mollis also showed a decreased leaf N content under 650 ppm CO<sub>2</sub> (Larigauderie et al., 1988). Declining N content under elevated CO<sub>2</sub> would thus mean a poorer tissue quality which could have a profound impact on herbivory, nutrient cycling and fertilizer use.

**Water use efficiency :** Higher atmospheric CO<sub>2</sub> concentration reduces stomatal aperture thereby reducing transpiration. This decrease in transpiration rate, together with the typical high-CO<sub>2</sub>-enhancement of net photosynthesis, accounts for the greater water use efficiency (ratio of carbon exchange rate to transpiration rate) in dry matter production under CO<sub>2</sub> enrichment (Warrick et al., 1984). Jones et al. (1984) observed that in soybean canopies grown under enriched-CO<sub>2</sub>, water use efficiency was enhanced 1.6 times than in ambient CO<sub>2</sub>, the absolute water requirements remaining the same in all treatments. Rogers et al. (1984) while experimenting with soybean (G. max) plants found that, transpiration per plant decreased with increasing CO<sub>2</sub> concentration inspite of increased leaf area per plant, leading to a decrease in water use per unit leaf area. B. mollis also showed an increased water use efficiency under

650 ppm CO<sub>2</sub> compared to 350 ppm CO<sub>2</sub> (Larigauderie et al., 1990).

Increased efficiency of water use alongwith decreased transpiration rates, could have far reaching effects on dry matter production in enhanced-CO<sub>2</sub> atmospheres especially under water-stressed conditions.

#### A.2. TREE SPECIES

Forests account for as much as two thirds of global photosynthesis (Kramer, 1981) and thus play a dominant role in the conversion of atmospheric CO<sub>2</sub> to fixed forms of carbon that have slow decomposition rates. The complexity of forest ecosystems and the technical challenges of quantifying their behaviour are few factors which have confined the number of studies on forest ecosystems. The studies available have been conducted with CO<sub>2</sub> enrichment of small trees and seedlings (Jarvis, 1989).

**Studies on Gymnosperms :** Growth chamber studies, on Douglas fir (Pseudotsuga menzeii) seedlings exposed to CO<sub>2</sub> (1000ppm) for 90 days, showed increased growth as a result of increased leaf photosynthesis (Purohit and Tregunna, 1976). Similar results have been obtained for seedling growth of lodgepole pine (Pinus contorta) and Sitka spruce (Picea sitchensis) (Canham and Mc Cavish, 1981). Under many

water and nutrient - stress conditions, seedling growth may be enhanced with elevated atmospheric CO<sub>2</sub> levels, [Pinus radiata, Pinus virginiana (Conroy et al., 1986, 1988)]. One open - top chamber experiment on long term CO<sub>2</sub> exposure was conducted with sapling of ponderosa pine (Pinus ponderosa) for 2.5 years (Surano et al., 1986). This pilot study showed that tree growth was enhanced upto a CO<sub>2</sub> level of 500 ppm but at 650 ppm growth was inhibited, an effect attributed to heat stress.

**Studies on Angiosperms :** A similar range of responses has been identified for seedlings of angiosperm deciduous species exposed to CO<sub>2</sub> enrichment [Quercus alba (Norby et al., 1986a), Liriodendron tulipifera (O'Neill, 1987)]. Long term growth responses of forest species to CO<sub>2</sub> enrichment remain speculative.

In the nutrient-cycling dynamics of forests, litter quality is another factor that could change with CO<sub>2</sub> enrichment. Litter produced at high CO<sub>2</sub> was predicted to be carbon rich and nitrogen poor (Norby et al., 1986b), leading to slower rates of decomposition.

Responses of mature trees to CO<sub>2</sub> enrichment have been evaluated using tree ring chronologies (Kienast and Luxmoore, 1988). Trees in temperate zones form distinct

annual growth rings. Since tree growth responds to CO<sub>2</sub> enrichment, it is expected that the historical change in CO<sub>2</sub> is recorded in tree-ring chronologies. Findings from modern tree-ring records indicate increases in growth that correlate with the increase in atmospheric CO<sub>2</sub> in recent decades (Kienast and Luxmoore, 1988).

Forest ecosystems thus need greater attention as the studies that have been carried out are restricted to few species specifically of Pinaceae family. The information gathered is insufficient to predict the responses of forest ecosystems to elevated CO<sub>2</sub> levels.

### A.3. AQUATIC SPECIES

Aquatic plant species that have been studied for their responses under elevated CO<sub>2</sub> are Eichhornia crassipes (Spencer and Bowes, 1986, Idso et al., 1987) Nymphaea marliac (Allen et al., 1990), Vallisneria americana (Titus et al., 1990) and Azolla pinnata (Idso et al., 1987, Allen et al., 1988), which is a pteridophyte. Among these E. crassipes is the most well investigated species. In E. crassipes (Spencer and Bowes, 1986), leaf number, as also the leaf area per plant increased under elevated CO<sub>2</sub> (600 ppm). A 32% increase in dry matter production in high CO<sub>2</sub> plants over ambient plants was noted. Flower production

increased substantially at 600 ppm CO<sub>2</sub>. Net photosynthesis increased by 40%, but this was not maintained as plants acclimated to high CO<sub>2</sub> over a 4-week period. Rubisco activity was 40% less after 4 weeks in 600 ppm CO<sub>2</sub>. Dark respiration rates of leaves, reduced by about one third in enriched plants. Transpiration rate of 600 ppm plants declined over the course of experiment especially on adaxial leaf surfaces. After four weeks in elevated as compared to ambient CO<sub>2</sub>, soluble protein content was 49% less, chlorophyll 26% less, and starch content 40% greater. Net photosynthesis of N. marliac (Allen et al., 1990) in 640 ppm CO<sub>2</sub> under conditions of high light and high temperature was 60% greater than in ambient CO<sub>2</sub> treatment. In A. pinnata an aquatic fern (Allen et al., 1988), net photosynthesis was influenced by significant interactions between CO<sub>2</sub> level and short wave solar radiation as well as air temperature. Under the favorable conditions of high light intensity and high temperature, the net photosynthesis rate of Azolla under 640 ppm CO<sub>2</sub> was 70% greater than for those in ambient CO<sub>2</sub> treatment. In V. americana, at pH 5 biomass increased 2.8 times at elevated CO<sub>2</sub> (770 ppm) compared to ambient (Titus et al., 1990). The information on response of aquatic plants to elevated CO<sub>2</sub> is limited. It is thus extremely difficult to assess their responses under CO<sub>2</sub> enrichment.

TABLE 1: A COMPARISON OF C<sub>3</sub> AND C<sub>4</sub> PLANT RESPONSE TO THE DOUBLING OF CO<sub>2</sub>  
(BASED ON LITERATURE SURVEY FROM 1968-1990)

PARAMETER	RESPONSE UNDER ELEVATED CO <sub>2</sub>	
	C <sub>3</sub>	C <sub>4</sub>
<b>CARBON EXCHANGE</b>		
Stomatal conductance	-	-
Net photosynthesis	+	0
Plant respiration	-	?
Decomposition of dead shoots	-	?
<b>GROWTH</b>		
Leaf area	+	+
Biomass	+	0
Photosynthate partitioning	M	?
Root/shoot ratio	+	0
<b>TISSUE COMPOSITION</b>		
Tissue N concentration	-	0
C/N ratio	+	0
Starch content	+	0

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**DEVELOPMENT/REPRODUCTION**

Tillering	+	?
Flowering time	M	?
Number of seeds	?	?

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**WATER USE**

Transpiration	-	-
Leaf water potential	+	+
Water use efficiency	+	+

---

+ Increase

- Decrease

M Mixed response

0 No response

? No information

A comparison of C<sub>3</sub> and C<sub>4</sub> plant response to the doubling of CO<sub>2</sub> levels (Table 1) shows that in relation to stomatal conductance, and transpiration, both plant groups show a decreasing trend. But with respect to leaf area, leaf water potential and water use efficiency, both C<sub>3</sub> and C<sub>4</sub> show an increasing trend. There are other parameters where a clear dichotomy in response of C<sub>3</sub> and C<sub>4</sub> has been observed. Net photosynthesis increases under enhanced CO<sub>2</sub> in C<sub>3</sub> while C<sub>4</sub> plants do not respond. Biomass and starch content increases in C<sub>3</sub> but not in C<sub>4</sub>. Tissue N concentration decreases in C<sub>3</sub> but is not affected in C<sub>4</sub> species. Many aspects like photosynthate partitioning, respiration, flowering time, seed number, decomposition rates under elevated CO<sub>2</sub> have not been studied well in case of C<sub>4</sub> species. Thus a comprehensive study of effect of elevated CO<sub>2</sub> on growth determining developmental, physiological and biochemical parameters of plants is required to predict their responses.

#### **Enhanced CO<sub>2</sub> in Relation to Environmental Factors**

Interactions between the atmospheric CO<sub>2</sub> and other growth limiting environmental variables and their effects on plant growth are complex and not studied well. Experimental studies indicate that elevated CO<sub>2</sub> concentration can reduce the deleterious impacts on growth, because of water -



shortages, low light intensity, temperature extremes or certain mineral deficiencies, notably nitrogen deficiency.

**Temperature** : A global warming of about 3°C is predicted for the year 2030 A.D. due to increasing levels of greenhouse gases in atmosphere (Jager, 1986). In this context, plant response to enriched CO<sub>2</sub> under higher temperature is important. It appears that in general the positive effect of higher CO<sub>2</sub> in stimulating photosynthesis is increased with higher temperature. Growth of C<sub>3</sub> plants is expected to increase by as much as 56% with a rise in surface air temperature of 3°C and atmospheric concentration of 640 ppm CO<sub>2</sub> (Cure and Acock, 1986). However, the effect of increasing temperature on the kinetic properties of rubisco, and solubility of CO<sub>2</sub> (relative to O<sub>2</sub>), which declines, could cause a negative feedback (Jordan and Ogren, 1984). Also temperature is important in determining the rate of growth of metabolic sinks (such as developing fruits) and high temperature adversely affect sink growth. All these factors would thereby feed back onto leaf photosynthesis and modulate the CO<sub>2</sub> response.

At low temperatures too, high CO<sub>2</sub> has been shown to have positive effects, by reducing the minimum temperature at which a plant grows and completes its life cycle (Sionit et al., 1981b). Okra plants (Abelmoschos

esculentor) were unable to complete their life-cycle in normal CO<sub>2</sub> at temperature below 23° (day)/17°C (night), while plants grown in 1000 ppm CO<sub>2</sub> at 20/14°C matured and produced fruit (Sionit et al., 1981). Thus with increasing atmospheric CO<sub>2</sub> levels the cultivation of okra may spread into cooler areas. Interaction of air temperature and elevated CO<sub>2</sub> would thus play an important role in determining growth and reproductive success of a species in the altered environment.

**Water** : Elevated CO<sub>2</sub> levels decrease transpiration rates and stomatal conductance of plants hence increasing their water use efficiency (Warrick et al., 1984). Wheat plants growing in elevated CO<sub>2</sub> under water stress (Sionit et al., 1981c) have been shown to osmoregulate more effectively. In wheat, Gifford (1979) reported that under extreme aridity, there was relative enhancement of yield due to CO<sub>2</sub> enrichment, because it allowed some grain growth where none occurred without extra CO<sub>2</sub>. Water-stressed soybean plants (Rogers et al., 1984) showed greater leaf tissue damage, lower leaf water potential, and higher stomatal resistance in low CO<sub>2</sub> than in high CO<sub>2</sub> grown plants. Starch accumulated in water stressed leaves of plants grown in CO<sub>2</sub> enriched environment. Under water stress coupled with elevated CO<sub>2</sub>, there was a significant increase in assimilate partitioning to roots in wheat (Gifford, 1979). This could be an

important response under field conditions if it were to allow roots to probe deeper layers of moist soil under CO<sub>2</sub> enriched conditions (Gifford, 1979).

Under growth-limiting water supply, growth of C<sub>3</sub> crops responds to higher CO<sub>2</sub> because of both photosynthetic and stomatal effects (Gifford, 1979) while growth of C<sub>4</sub> species responds because of stomatal effects alone. Thus for both C<sub>3</sub> and C<sub>4</sub> species, the less the availability of water, the greater the 'relative enhancement' of growth by high CO<sub>2</sub> concentrations (Gifford 1979).

### Nutrients

**Nitrogen** : Low nitrogen supply reduces growth of all species, under ambient CO<sub>2</sub>. With doubling of CO<sub>2</sub> concentration, however, C<sub>3</sub> non leguminous plant species will still register a relative enhancement in dry weight growth even under nitrogen stress. The weight of cotton plants almost doubled, both under 2mM or 24mM nitrate in the nutrient solution when CO<sub>2</sub> concentration was increased from 330 to 640ppm, whereas for corn, a C<sub>4</sub> non legume, the increase was only 20% (Wong, 1979). In wheat, Sionit et al. (1981) found that the stimulation of dry-matter accumulation by 675 ppm plants compared to 350 ppm CO<sub>2</sub> plants increased with increasing nutrient availability. In legumes such as

soybeans or peas, high CO<sub>2</sub> leads to greater biological nitrogen fixation which could be attributed to the production of more nodules on an elaborate root system, rather than to greater specific activity of nodules (Finn and Brun, 1982). Under CO<sub>2</sub> enriched conditions, N-use efficiency of a plant tends to increase. The increased efficiency could be due to a reduced investment in photosynthetic machinery (which has a high N-requirement) per unit of photosynthetic assimilate produced (Sage et al., 1989).

**Phosphorus :** P-deficient plants of Z. mays, S. bicolor and G. max (Pettigrew et al., 1990) under CO<sub>2</sub> saturating condition had lower net photosynthetic rates than P-sufficient plants. This could be because inorganic phosphate (Pi) plays an important role in regulating transport of triose - phosphate sugars out of chloroplasts via the phosphate translocator; in P-deficient plants this mechanism being disrupted could have lead to end-product inhibition of photosynthesis (Pettigrew et al., 1990). On the contrary P-deficient bean (Vicia faba) plants have been found to be even more responsive to high CO<sub>2</sub> than were plants grown with adequate P (Goudriaan and de Ruiter, 1983). Further studies are required to elucidate the actual mechanism of interaction of high CO<sub>2</sub> and P.

**Potassium** : There is little information on interaction of potassium with atmospheric CO<sub>2</sub> enrichment on plants. In potato, Goudriann and de Ruiten (1983) noted negative effect of increased CO<sub>2</sub>.

**Sodium** : Sodium is an essential element for C<sub>4</sub> photosynthesis. The signs of sodium deficiency in the C<sub>4</sub> species, Amaranthus tricolor and Atriplex spongiosa were alleviated when the species were grown in conditions of high CO<sub>2</sub> concentration (1500 ppm) (Johnston et al., 1984). Sodium sufficient C<sub>4</sub> plants were relatively unaffected by the CO<sub>2</sub> treatments (Johnston et al., 1984). Schwarz and Gale (1984), on the other hand have shown that tolerance of saline (excess sodium) conditions is increased by CO<sub>2</sub> enrichment to 2500 ppm. This effect was ascribed to, a shortage of photosynthate in salt stressed plants, made up by enhanced CO<sub>2</sub>, or to reduced demand for saline water because of CO<sub>2</sub> reduced transpiration under enhanced CO<sub>2</sub>.

**Light** : CO<sub>2</sub> enrichment increases crop growth and yield at low light intensities which are otherwise growth limiting under conditions of ambient CO<sub>2</sub>. The relative enhancement of growth can even be greater than at high light level, as has been found for wheat (Gifford, 1979). The mechanism of growth response to CO<sub>2</sub> depends on two factors under photosynthetically limiting light intensities. One is

that the quantum yield of leaf photosynthesis close to light compensation point (the light intensity at which  $\text{CO}_2$  uptake by a leaf is just balanced by respiratory release of  $\text{CO}_2$ ) is  $\text{CO}_2$  dependent in  $\text{C}_3$  species, but not in  $\text{C}_4$  species (Ehleringer and Bjorkman, 1977). If the whole plant respiration is less under high  $\text{CO}_2$ , then the light compensation point is lowered and some growth is achieved at light intensities that otherwise would prove insufficient for photosynthesis, which explains the pattern shown by wheat under high  $\text{CO}_2$  and low intensity (Gifford, 1979). In case of soybean, which has shown increased respiration under high  $\text{CO}_2$ , the relative enhancement of growth by high  $\text{CO}_2$  appears equal at low and high light intensities (Sionit et al., 1982).

#### B. ECOSYSTEM LEVEL RESPONSES

The knowledge about effects of  $\text{CO}_2$  at community and ecosystem level is very limited. Recently, the following two natural ecosystems have been studied at Toolik lake, Alaska and Chesapeake Bay, to gain some insight into ecosystem functioning in response to  $\text{CO}_2$  enrichment.

- (1) Moist tussock tundra at Toolik lake in the foothills of the Brooks Range in Alaska (Oechel and Riechers, 1986, Tissue and Oechel, 1987). Temperature controlled greenhouses were used to maintain elevated  $\text{CO}_2$  levels

(510 and 680 ppm). The arctic tundra ecosystem was floristically diverse and comprised of C<sub>3</sub> species. Dominant plant species was cotton grass, Eriophorum vaginatum.

- (2) Coastal salt-marsh on the Chesapeake Bay (Curtis et al., 1989a, 1989b, 1990, Drake et al., 1989, Ziska et al., 1990). Here open-top chambers were used to create test atmospheres of normal ambient & elevated CO<sub>2</sub> (normal ambient + 340ppm). The coastal marsh system was comprised of two higher plants, S. olneyi (C<sub>3</sub>) and S. patens (C<sub>4</sub>), both often occurring in monospecific stands, and a mixed community of S. olneyi, S. patens and D. spicata (C<sub>4</sub>).

Significant ecosystem level effects were noted in both the arctic and salt marsh.

**Net carbon storage** : In arctic ecosystem short-term exposure to elevated CO<sub>2</sub> resulted in immediate positive ecosystem carbon gain while ambient CO<sub>2</sub> chambers achieved it 6 d later. Over a 74 d growing season the tussock tundra under ambient CO<sub>2</sub> had a net carbon loss (-53.4 g C m<sup>-2</sup> y<sup>-1</sup>), whereas elevated CO<sub>2</sub> chambers showed net carbon acquisition (206.5 g C m<sup>-2</sup> y<sup>-1</sup>). Homeostatic adjustment of whole ecosystem carbon flux was complete within three years

(Grulke et al., 1990). The marsh increased carbon storage under elevated CO<sub>2</sub> but no changes in nutrient relations were observed. The C<sub>4</sub> S. patens stands showed results similar to S. olneyi, except there was no increase in ecosystem carbon storage (Ziska et al., 1990).

**Biomass :** In tussock tundra root biomass and root : shoot ratio generally decreased at elevated CO<sub>2</sub> (Tissue and Oechel, 1987). Growth under elevated CO<sub>2</sub> resulted in an 83% increase in root dry mass in Scirpus community. S. patens community and C<sub>4</sub> component of mixed community showed no increase in root growth under elevated CO<sub>2</sub> (Curtis et al, 1990).

**Dark respiration :** Elevated CO<sub>2</sub> concentration had no significant effects on tundra ecosystem dark respiration rates (Grulke et al., 1990). Net ecosystem respiration decreased in salt marsh system (Curtis et al., 1989a).

**Nitrogen content:** In the arctic tundra, elevated CO<sub>2</sub> tended to decrease nitrogen content and increase C/N ratio (Tissue and Oechel, 1987). Nitrogen content (%) of roots of S. olneyi was lower under elevated CO<sub>2</sub> compared to ambient grown plants. No effect on nitrogen content was observed in S. patens or D. spicata (Curtis et al., 1990).



**Water relations:** There was little long term effect on evapotranspiration, or water-use (Tissue and Oechel, 1987) in tussock tundra system. In coastal marsh system stands of C<sub>3</sub> S. olneyi, showed improvements in water relations under elevated CO<sub>2</sub>. Evapotranspiration decreased by 30% in both C<sub>3</sub> and C<sub>4</sub> stands in the salt marsh system, resulting in 80-100% increase in water use efficiency.

A list of species (Table 2) and families (Table 3) studied for their response to CO<sub>2</sub> doubling shows that the study is spread over fifty four species belonging to eighteen families. Table 2 clearly indicates that a greater emphasis has been placed on response of crop plants (39% of studies) and their associated weeds (20% of studies) to elevated CO<sub>2</sub>. However, in relation to uncultivated herbaceous species, tree species as well as aquatic species not many studies have been done. Leguminosae is the most well studied family among angiosperms while among gymnosperms studies have been limited to Pinaceae family and specifically to genus Pinus (Table 3). Among trees only short term experiments using tree seedlings have been done. Only one long term study (2.5 yr) using saplings of Pinus ponderosa (Surano et al., 1986).

Information on aquatic plants is limited to four species only, one of them being a fern, Azolla pinnata.

TABLE 2 : LIST OF PLANT SPECIES STUDIED FOR ELEVATED CO<sub>2</sub> RESPONSE  
(BASED ON LITERATURE SURVEY FROM 1968 TO 1990)

Plant Species	Meta- bolism	Method of Study*	Duration of Study**	Cate- gory***	Family	Reference
<b>HERBACEOUS SPECIES</b>						
<u>Abelmoschus esculentus</u>	C <sub>3</sub>	GC	L	C	Malvaceae	Sionit et al., 1981.
<u>Abutilon theophrasti</u>	C <sub>3</sub>	GC	S,L	UC	Malvaceae	Patterson and Flint, 1980, Garbutt et al., 1990.
<u>Amaranthus tricolor</u>	C <sub>4</sub>	GC	L	UC	Amaranthaceae	Johnston et al., 1984
<u>Amaranthus retroflexus</u>	C <sub>4</sub>	GC	S	UC	Amaranthaceae	Garbutt et al., 1990
<u>Ambrosia artemisiifolia</u>	C <sub>3</sub>	GC	S	UC		Garbutt et al., 1990
<u>Atriplex hastata</u>	C <sub>3</sub>	GC	L	UC	Chenopodiaceae	Johnston et al., 1984
<u>Atriplex spongiosa</u>	C <sub>4</sub>	GC	L	UC	Chenopodiaceae	Johnston et al., 1984
<u>Beta vulgaris</u>	C <sub>3</sub>	GC	S	C	Chenopodiaceae	Wyse, 1980
<u>Brassica oleracea</u>	C <sub>3</sub>	GC	L	C	Cruciferae	Sage et al., 1989
<u>Bromus mollis</u>	C <sub>3</sub>	GC	L	UC	Gramineae	Larigauderie et al., 1988
<u>Cassia obtusifolia</u>	C <sub>3</sub>	GC	S,L	UC	Leguminosae	Pattern son Flint, 1982
<u>Chenopodium album</u>	C <sub>3</sub>	GC	L	UC	Chenopodiaceae	Sage et al., 1989, Garbutt et al., 1990
<u>Crotalaria spectabilis</u>	C <sub>3</sub>	GC	S,L	UC	Leguminosae	Patterson and Flint, 1984
<u>Daucus carota</u>	C <sub>3</sub>	OTC	L	C	Umbelliferae	Idso et al., 1987
<u>Distichlis spicata</u>	C <sub>4</sub>	OTC	L	UC	Gramineae	Curtis et al., 1989, Curtis and Balduman, 1990.
<u>Eriophorum vaginatum</u>		GC	L	UC		Tissue and Dechel, 1987; Grulke et al., 1990
<u>Glycine max</u>	C <sub>3</sub>	GC	S,L	C	Leguminosae	Patterson and Flint, 1980; Clough et al., 1981; Finn and Brun, 1982; Patterson and Flint, 1982; Jones et al., 1984; Cure et al., 1987; Rogers et al., 1984, 1986
		OTC	L			

TREE SPECIES

<u>Acer macrophyllum</u>	C <sub>3</sub>	GC	S		Aceraceae	Bailey et al., 1991
<u>Liquidamber styraciflua</u>	C <sub>3</sub>	GC	S		Hamamelidaceae	Tolley and Strain, 1984; Fetcher et al., 1988
<u>Liriodendron tulipifera</u>	C <sub>3</sub>	GC	S		Magnoliaceae	O'Neill, 1987
<u>Picea sitchensis</u>	C <sub>3</sub>	GC	S		Pinaceae	Canham and Mc Cavish, 1981
<u>Pinus contorta</u>	C <sub>3</sub>	GC	S		Pinaceae	Canham and Ma Cavish, 1981
<u>P. ponderosa</u>	C <sub>3</sub>	GC	S,L		Pinaceae	Green and Wright, 1977; Surano et al., 1986
<u>P. radiata</u>	C <sub>3</sub>	GC	S		Pinaceae	Conroy et al., 1986,1988
<u>P. strobus</u>	C <sub>3</sub>	GC	S		Pinaceae	Funsh, 1970
<u>P. taeda</u>	C <sub>3</sub>	GC	S		Pinaceae	Tolley and Strain 1984; Fetcher et al., 1988
<u>P. virginiana</u>		GC	S		Pinaceae	Conroy et al., 1986, 1988
<u>Pseudotsuga menzeii</u>	C <sub>3</sub>	GC	S		Pinaceae	Purohit and Tregunna, 1976
<u>Quercus alba</u>	C <sub>3</sub>	GC	S		Fagaceae	Norby et al., 1986

AQUATIC SPECIES

<u>Azolla pinnata</u>	C <sub>3</sub>	OTC	L	UC	Salviniaceae	Idso, 87, Allen et al., 1988
<u>Eichhornia crassipes</u>	C <sub>3</sub>	GC OTC	L	UC	Pontederiaceae	Spencer and Bowes, 1986 Idso, et al., 1987
<u>Nymphaea marliac</u>	C <sub>3</sub>	OTC	L	UC	Nymphaeaceae	Allen et al., 1990
<u>Vallisneria americana</u>	C <sub>3</sub>	GC	L	UC	Hydrocharitaceae	Titus et al., 1990

\* GC - Growth Chamber/Environment Chamber/Greenhouse  
/Controlled Environment Rooms/Glasshouse/Plant Chamber  
OTC- Open top Chamber

\*\* S- Short term study  
L- Long term study

\*\*\* UC - Uncultivated  
C - Cultivated

TABLE 3 :LIST OF PLANT FAMILIES AND SPECIES STUDIED IN RELATION  
 TO CO<sub>2</sub> ENRICHMENT  
 (BASED ON LITERATURE SURVEY FROM 1968 TO 1990)

Family	No. of species studied
Leguminosae	9
Gramineae	8
Pinaceae	8
Chenopodiaceae	4
Solanaceae	4
Malvaceae	3
Amaranthaceae	2
Compositae	2
Cyperaceae	2
Umbelliferae	1
Aceraceae	1
Hamamelidaceae	1
Magnoliaceae	1
Fagaceae	1
Salviniaceae	1
Pontederiaceae	1
Nymphaeaceae	1
Hydrocharitaceae	1

TABLE 4 :STUDIES ON PLANT RESPONSES TO DIFFERENT CONCENTRATIONS  
 OF CO<sub>2</sub> RANGING FROM 300 - 1500 ppm  
 (BASED ON LITERATURE SURVEY FROM 1968 TO 1990)

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CO <sub>2</sub> CONCENTRATION RANGE (ppm)	NO. OF STUDIES
300 - 399	66
400 - 499	9
500 - 599	8
600 - 699	43
700 - 799	8
800 - 899	2
900 - 999	14
1000- 1099	14
1100- 1199	1
1200- 1299	1
1500	5

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This survey shows that database to comprehend and predict plant responses to elevated CO<sub>2</sub> is rather limited. Another aspect relates to the fact that in most of the species plant responses have been studied in relation to doubling of CO<sub>2</sub> (600-650 ppm). The impact of intermediate levels of CO<sub>2</sub> concentrations ranging from 400-500 ppm, likely to be encountered in next few years, is yet to be studied at individual and community level.

Species level studies provide valuable specific information but it is of little help in comprehending their response in a community under field conditions. The plant responses in a community or natural ecosystems are likely to be much different. Species composition in communities and natural ecosystems is likely to change depending on their relative competitive ability and reproductive success under altered CO<sub>2</sub> regime. Changes in mineral composition of plants under elevated CO<sub>2</sub> might play an important role in changing pest and herbivore preferences. Litter decomposition under high CO<sub>2</sub> could be slower due to a likely increase in C/N ratio. This would result in a slow nutrient release which may further complicate nutrient status of soil and affect plant response. Studies of intra- and inter-specific competition in relation to elevated CO<sub>2</sub> on various types of economically important and other wild

plants are important to anticipate responses of agro-ecosystems and natural ecosystems.

Detailed studies under elevated CO<sub>2</sub> at species and ecosystem level are required in relation to local variations of climatic, edaphic and anthropogenic factors. Such primary data on plants would provide a sound basis for assessing impact of elevated CO<sub>2</sub> at local and regional levels.

## PART II: EFFECT OF ENHANCED LEVELS OF CO<sub>2</sub> ON SPINACEA OLERACEA

### Materials and Methods:

**Exposure Chamber :** An open-top type of exposure chamber of 2.5 m height and 2 m diameter was constructed using locally available materials (Plate 1, 2) based on design given by Rogers et al., 1983. The chamber frame consisted of eight bamboo poles placed equidistant forming a circle of 2 m diameter. Poles were tied with one another with a thick cotton tape woven around them at a height of 0.5 m and 1.5 m from the ground to provide support and stability. This prevented the caving in of the chamber due to strong winds or under its own weight. The chamber was covered with transparent PVC plastic sheet. An entry point to the chamber was provided by leaving the plastic sheets unsealed. The entry was kept covered by overlapping plastic flaps when not in use.

Inside the chamber, at a height of 0.3 m from the ground, an inflatable plastic air-delievery tube of 0.114 m diameter was fixed along the inner wall of the exposure chamber. Equally placed holes (0.5 cm diameter) were punched on the inside of the air delievery tube in such a manner that the flow of CO<sub>2</sub> enriched air into the chamber keeps it fully inflated, and distributes it uniformly inside the chamber.





Plate 1 Open top chamber used in the experiment

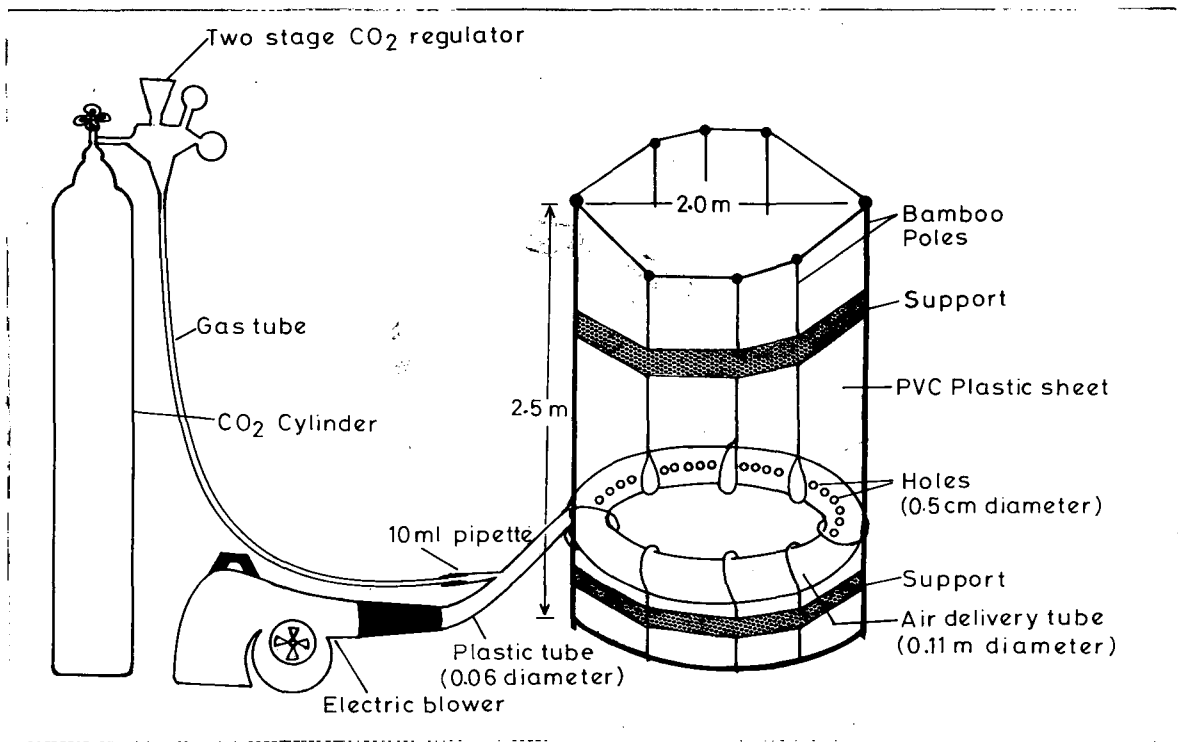


Plate 2 Schematic diagram of open top chamber system used in the experiment

**CO<sub>2</sub> supply :** The source of carbon dioxide was compressed CO<sub>2</sub> cylinders ( 27 kg) fitted with a double stage CO<sub>2</sub> regulator. The gas cylinder was connected to an electric blower (flow rate = 1.5 l/min). The blower was connected to the air delivery tube by a thick plastic pipe (diameter = 0.06m). A 10 ml pipette, connected with help of a gas tube to the CO<sub>2</sub> cylinder, released CO<sub>2</sub> gas into the plastic pipe connected to blower. This ensured through mixing of air and CO<sub>2</sub> which was finally released into the air delivery tube. The open top of the chamber acted as the outlet.

CO<sub>2</sub> levels inside the exposure chamber were monitored every three hr with the help of a Portable Photosynthetic System (LICOR LI-6000), which has an in built IR analyzer. CO<sub>2</sub> levels inside the chamber were maintained at 600 ± 50 ppm.

An identical open top chamber was constructed to keep control plants, the only difference being that the air supplied to this chamber was not enriched with CO<sub>2</sub>. The chambers were constructed in an open, unshaded area in the ecology lab garden.

**Plant Materials :** The following two cultivars of Spinacea oleracea (C<sub>3</sub>) were chosen for the experimental

study:

- i cv. 'All green'
- ii cv. 'Banerjee's giant'

Thirty earthen pots of 15 cm height, 20 cm diameter were filled with well manured garden soil. Plants of both varieties were raised from seeds in labelled pots. Eighteen days after planting, ten pots of each variety having similar size plants were chosen. Thinning was done and only 3 plants per pot were maintained. Out of the ten pots of each variety, five were kept in CO<sub>2</sub> - enriched chamber and rest five were kept in the chamber supplied with ambient air only. The plants in experimental pots were properly labelled and were continuously exposed to elevated CO<sub>2</sub> for ten days from 19.4.91 to 29.4.91. The plants were watered regularly, and 0.1 % aqueous solution of malathion was sprayed once in five days as a prophylactic measure to protect the experimental plants from pests.

**Leaf Area :** Length (l) and breadth (b) of each leaf was measured and leaf area was calculated by  $l \times b \times$  multiplying factor. The multiplying factor was calculated as follows. Fifty leaves of Spinacea oleracea were taken and their length (l) and breadth (b) was recorded.  $l \times b$  gave apparent leaf area. The actual leaf area was determined with the help of graph paper. Actual leaf area divided by

apparent leaf area gave the multiplying factor, which in this case was 0.660. Leaf area of each plant before the experiment and after terminating the experiment was estimated.

**Chlorophyll Estimation:** 0.5 g of fresh leaf tissue was homogenized in 10 ml of 80% acetone. The homogenate was centrifuged at 2000 g for 5 minutes. The clear green supernatant was taken and kept in a tube covered with aluminium foil. The pellet was re-extracted with another 10 ml of 80% acetone, centrifuged again and the supernatants from both steps were pooled together and the final volume made to 25 ml. The absorbance was measured at 645 and 663 nm (for chlorophyll estimation), at 480 and 510 nm (for carotene estimation) using Spectronic - 20. Chlorophyll a, b and carotene content was calculated according to following formulae (in mg/0.5g)

$$\text{Chlorophyll (a)} = \frac{12.7A_{663} - 2.69A_{645}}{a \times 1000 \times w}$$

$$\text{Chlorophyll (b)} = \frac{22.9A_{645} - 4.68A_{663}}{a \times 1000 \times w}$$

$$\text{Total Chlorophyll (mg/o.5g)} = \frac{20.2A_{645} + 8.02A_{663}}{a \times 1000 \times w} \times v$$

$$\text{Carotene} = \frac{7.6A_{480} - 1.49A_{510}}{a \times 1000 \times w} \times v$$

All values in mg/0.5 g leaf tissue were converted to mg/g.

- A = Absorbance at that particular wavelength  
a = Length of light path in the cell (usually 1 cm)  
v = Volume of sample  
w = weight of leaf tissue taken.

**Biomass:** After 10 days of exposure plants were harvested along with the root system. Each plant was carefully washed to remove the soil particles with a fine brush. Root and shoot were separated and put in pre weighed labelled beakers. For dry weight determination plant material was kept in an electric oven at 85° C for 48 hr and root : shoot ratios were calculated.

**Starch Content:** Starch content was determined using a simpler version of Pucher's method (Pucher et al., 1948). 250 mg of dry ground plant material was taken in a test tube. To this 200 mg of fine sand and 5 ml distilled water was added and mixed. The tube was heated in a boiling waterbath for 15 min to gel the starch. After cooling the

test tube 5 ml 60% HClO<sub>4</sub> was added whilst mixing. The tissue was grounded against the side of test tube for 20 min then transferred to a 100 ml volumetric flask and diluted to volume. Allowed to settle. An aliquot of 5 ml was transferred to a 50 ml volumetric flask. To this few drops of indicator solution (0.1% Phenol red, in 90% ethyl alcohol) were added, then NaOH (M) was added until solution turns red. Acetic acid (10% v/v) was added to destroy colour and then added further 2.5 ml. To this 0.5 ml KI solution (10% w/v), 5.0 ml KIO<sub>3</sub> solution (0.0125 M) were added. Shook well and diluted to volume. Absorbance was measured at 680 nm. A calibration curve was prepared using a range of standard starch solutions, and used to obtain mg starch in the sample aliquot. Percentage of starch content was calculated according to the following formula:

If C = mg starch obtained from calibration curve then,

$$\text{Starch (\%)} = \frac{C(\text{mg}) \times \text{solution volume (ml)}}{10 \times \text{aliquot (ml)} \times \text{sample wt (g)}}$$

## Results

Effect of elevated CO<sub>2</sub> (600 ± 50 ppm) on leaf area in Spinacea oleracea : (Table 5, Fig 1,2) Average leaf area per plant registered an increase under elevated CO<sub>2</sub> in both cultivars after ten days of exposure continuously. In S. oleracea cv. All Green average leaf area per plant (ambient CO<sub>2</sub>) was 30.21 cm<sup>2</sup> and 58.68 cm<sup>2</sup> (elevated CO<sub>2</sub>) amounting to a 94.2% increase over control. In S. oleracea cv. Banerjee's Giant average leaf area per plant was 26.36 cm<sup>2</sup> (ambient CO<sub>2</sub>) and 36.94 cm<sup>2</sup> (elevated CO<sub>2</sub>) resulting in 40.14% increase over control.

Effect of elevated CO<sub>2</sub> (600±50 ppm) on chlorophyll content in S. oleracea : (Table 6, Fig 3,4). There was a reduction in chlorophyll and carotene content in both cultivars under elevated CO<sub>2</sub>. In S. oleracea cv. All Green a 23.97% reduction in Chl (a), 39.47% in Chl (b) over control was observed. Total chlorophyll decreased from 1.005 mg/g leaf tissue (ambient CO<sub>2</sub>) to 0.706 mg/g (elevated CO<sub>2</sub>), a 29.75% reduction. Carotene content decreased to 34.85% over control. Chl a/b ratio increased from (ambient CO<sub>2</sub>) to 2.11 (elevated CO<sub>2</sub>). In S. oleracea cv. Banerjee's Giant Chl (a) did not show any reduction whereas Chl (b) decreased by 23.2% over control. Total Chl decreased from 0.732 mg/g (ambient CO<sub>2</sub>) to 0.681 (elevated CO<sub>2</sub>), amounting



to 6.97% reduction. Carotene content decreased by 8.53%. Chl a/b ratio increased from 2.3 to 3.05 under elevated CO<sub>2</sub>.

**Effect of elevated CO<sub>2</sub> (600 ± 50 ppm) on biomass in S. oleracea :** (Table 7, Fig 5,6) In S. oleracea cv. All Green root biomass showed a 296% increase whereas shoot biomass a 83.5% increase over control. Root/Shoot ratio increased from 0.161 (ambient CO<sub>2</sub>) to 0.348 (elevated CO<sub>2</sub>). In S. oleracea cv. Banerjee's Giant root biomass showed a 188.8% increase whereas shoot biomass a 73.5% increase over control. Root/Shoot ratio increased from 0.096 (ambient CO<sub>2</sub>) to 0.160 (elevated CO<sub>2</sub>).

**Effect of elevated CO<sub>2</sub> on foliar starch content in Spinacea oleracea :** (Table 8, Fig 7) Foliar starch content increased from 2.3% (ambient CO<sub>2</sub>) to 4.0% (elevated CO<sub>2</sub>) in S. oleracea cv. All Green. In S. oleracea cv. Banerjee's Giant it increased from 1.2% (ambient CO<sub>2</sub>) to 2.3% (elevated CO<sub>2</sub>).

TABLE 5: EFFECT OF ELEVATED CO<sub>2</sub> (600 ±50 ppm) ON LEAF AREA IN SPINACEA OLERACEA

Sample	Period of Exposure (days)	Average Leaf Area Plant <sup>-1</sup> (cm <sup>2</sup> )		Increase (%) in Leaf Area over Control
		0	10	
S. oleracea (cv. All Green)	Control	21.48	30.21	
	Elevated CO <sub>2</sub>	23.91	58.68	94.2
S. oleracea (cv. Banerjee's Giant )	Control	16.34	26.36	
	Elevated CO <sub>2</sub>	16.29	36.94	40.14

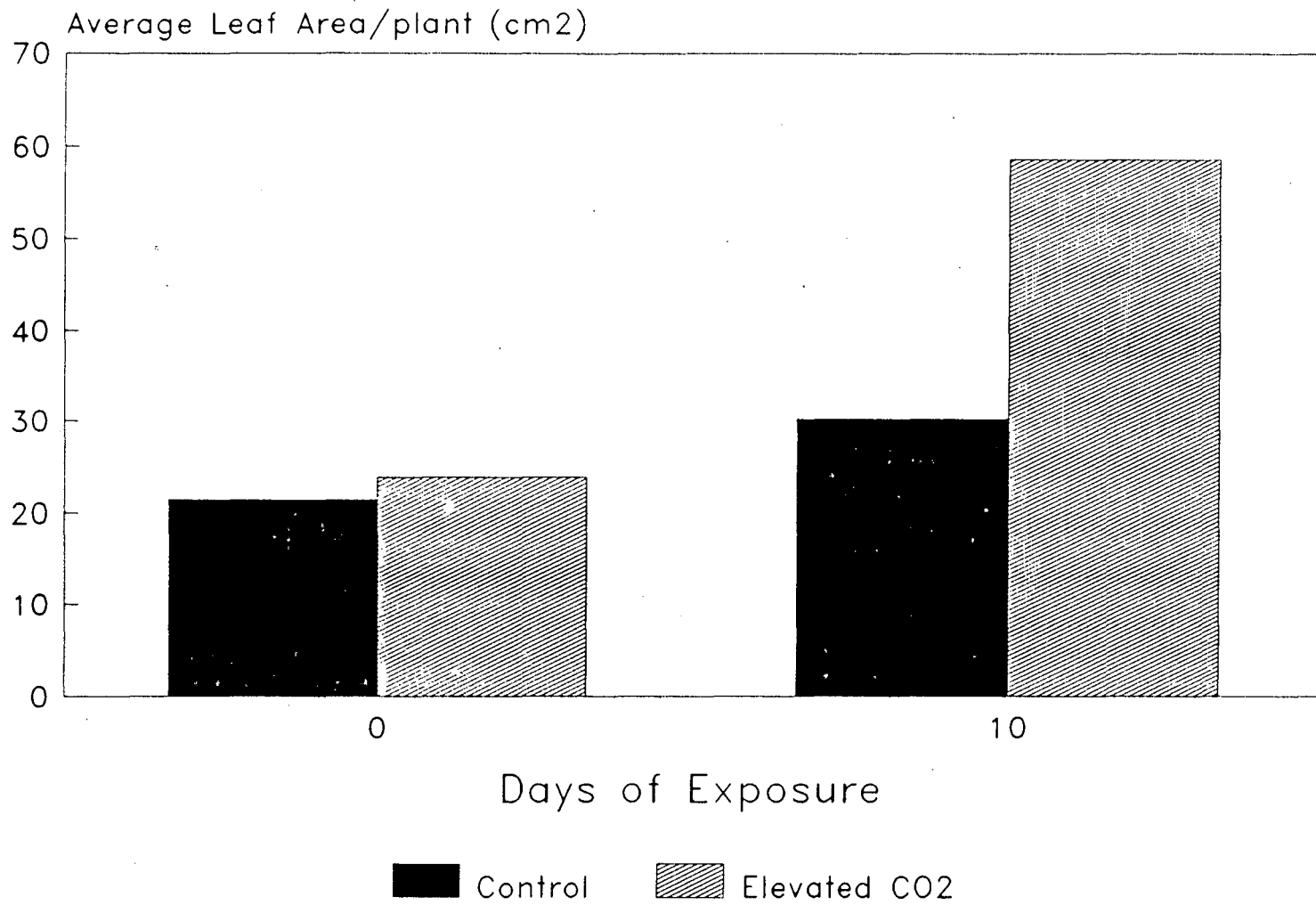


Fig 1 Increase in leaf area (*S oleracea* cv. All green) after ten days exposure to elevated CO<sub>2</sub> (600 ± 50 ppm)

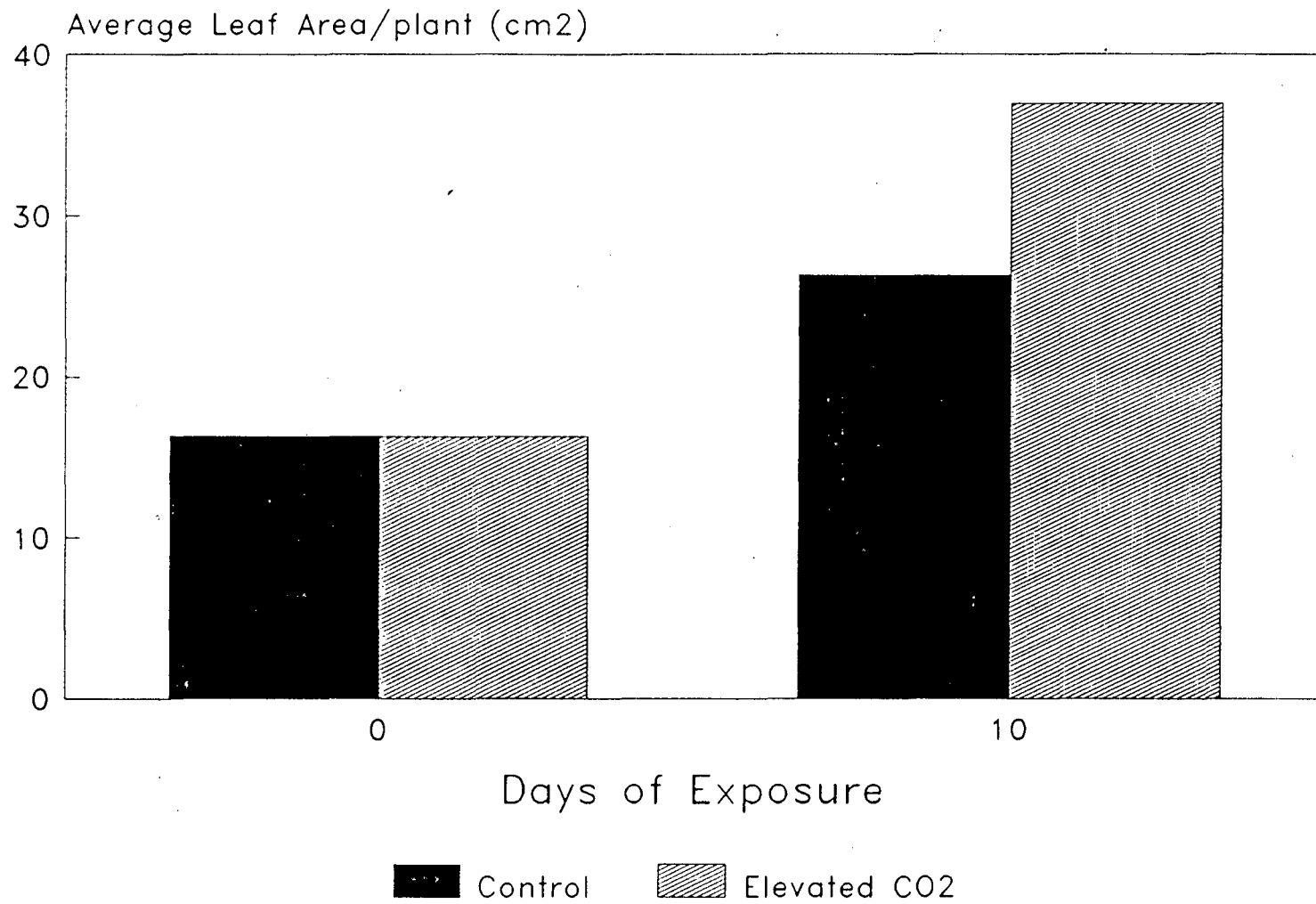


Fig 2 Increase in leaf area (*S. oleracea* cv. Banerjee's Giant) after ten days exposure to elevated CO<sub>2</sub> (600 ± 50 ppm)

TABLE 6: EFFECT OF ELEVATED CO<sub>2</sub> (600 ±50 ppm) ON CHLOROPHYLL CONTENT IN SPINACEA OLERACEA

Sample	Chl(a)* Reduction (%)over control	Chl(b)* Reduction (%)over control	Total Chl* Reduction (%)over control	Carotene* Reduction (%)over control	Chl a/b ratio	
S. oleracea (cv.All Green)	Control	0.630	0.375	1.005	0.396	1.68
	Elevated CO <sub>2</sub>	0.479 23.97	0.227 39.47	0.706 29.75	0.258 34.85	2.11
S. oleracea (cv.Banerjee's Giant)	Control	0.513	0.219	0.732	0.293	2.3
	Elevated CO <sub>2</sub>	0.513 0.00	0.168 23.2	0.681 6.97	0.268 8.53	3.05

\* Value expressed in mg/g leaf tissue

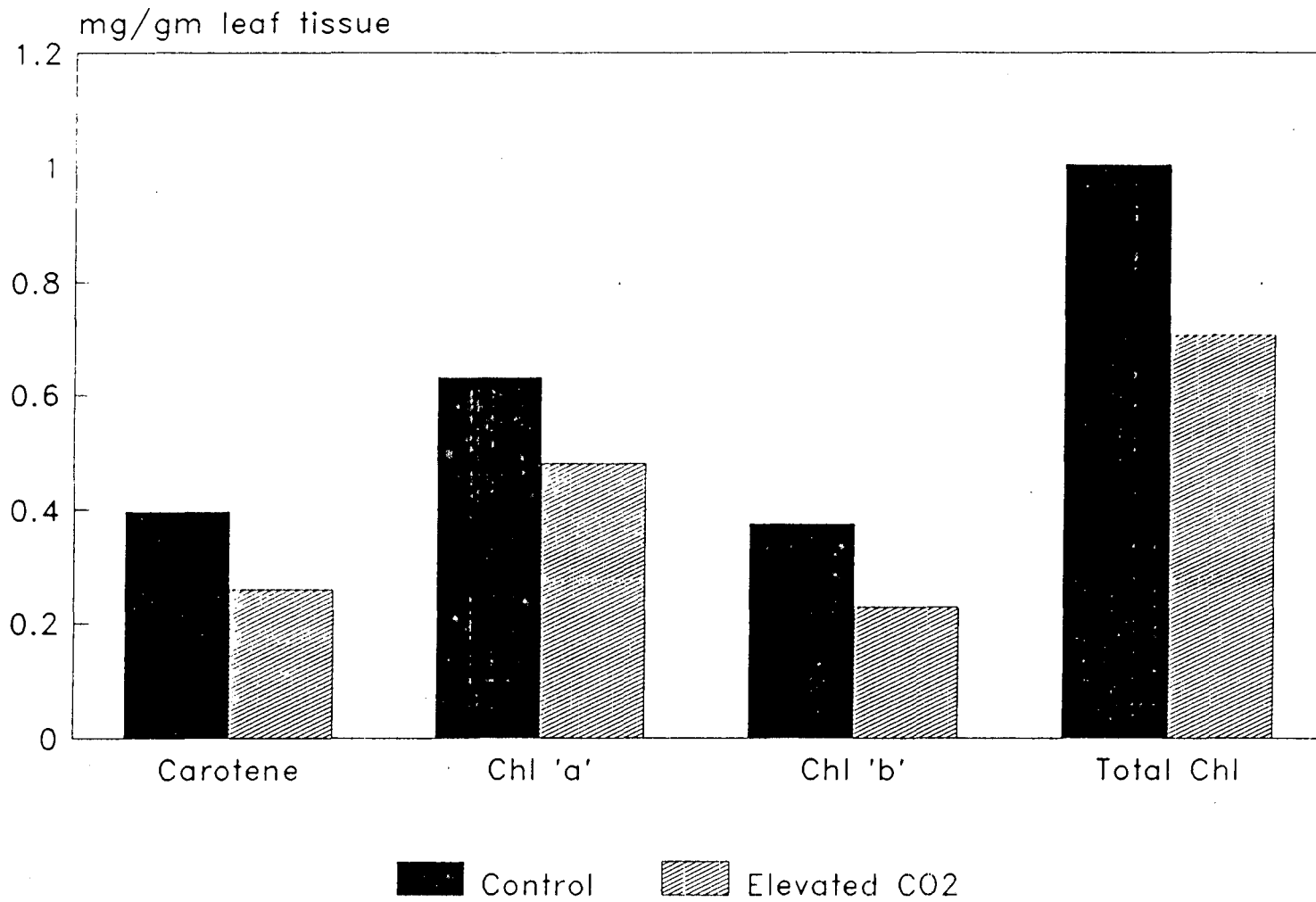


Fig 3 Decrease in chl (a), chl (b), total chl and carotene (*S. oleracea* cv. All Green) after ten days exposure to elevated CO<sub>2</sub> (600 ± 50 ppm)

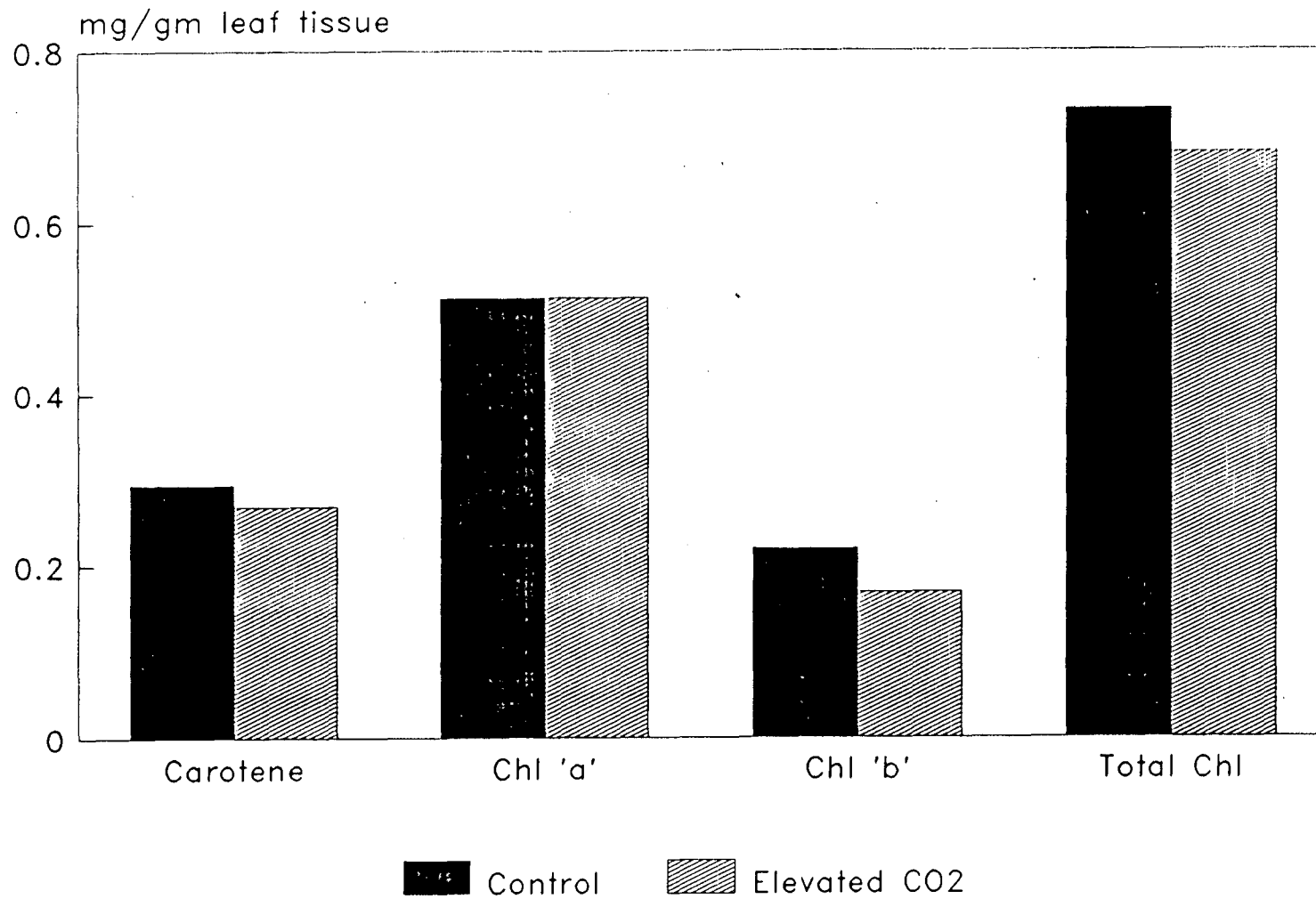


Fig 4 Decrease in chl (a), chl (b), total chl and carotene (*S. oleracea* cv. Banerjee's Giant) after ten days exposure to elevated CO<sub>2</sub> (600 ± 50 ppm)

TABLE 7: EFFECT OF ELEVATED CO<sub>2</sub> (600 ±50 ppm) ON BIOMASS IN SPINACEA OLERACEA

Sample		Root Biomass (g)	Increase (%) over control	Shoot Biomass (g)	Increase (%) over control	Root/Shoot
S. oleracea (cv. All Green)	Control	0.051		0.316		0.161
	Elevated CO <sub>2</sub>	0.202	296	0.580	83.5	0.348
S. oleracea (cv. Banerjee's Giant)	Control	0.009		0.094		0.096
	Elevated CO <sub>2</sub>	0.026	188.8	0.163	73.5	0.160



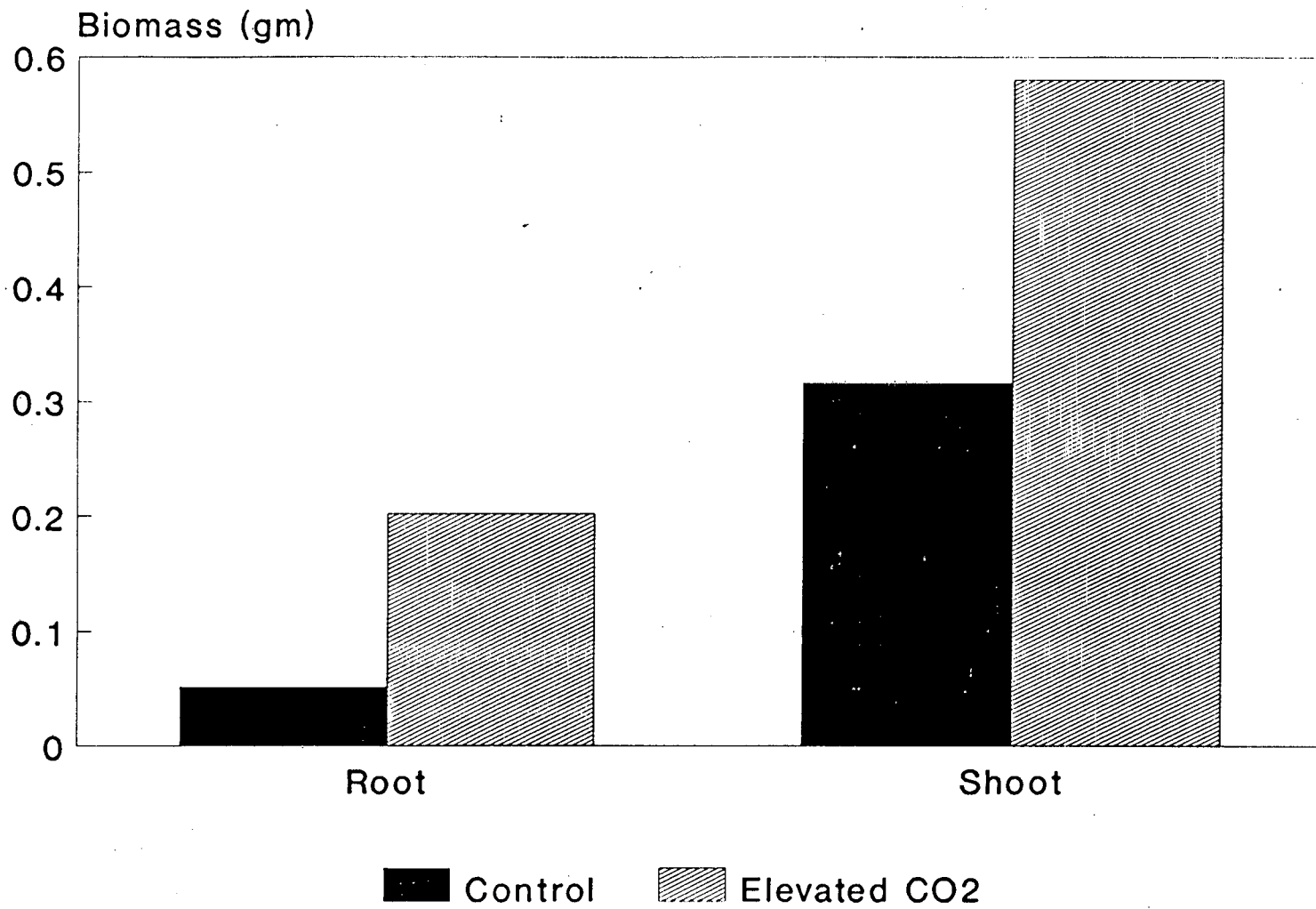


Fig 5 Increase in root and shoot biomass (*S. oleracea* cv. All Green) after ten days exposure to elevated CO<sub>2</sub> (600 ± 50 ppm)

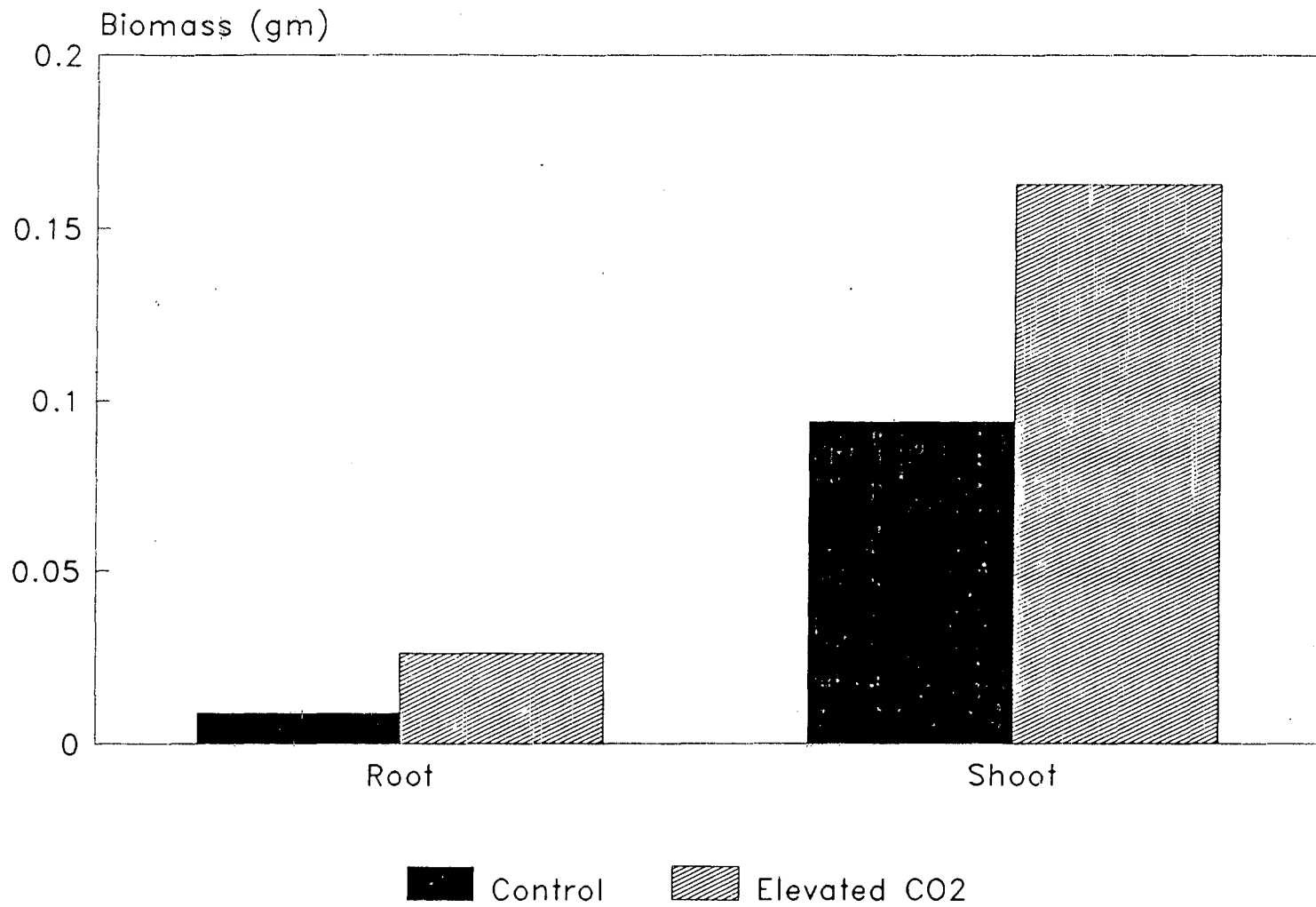


Fig 6 Increase in root and shoot biomass (*S. oleracea* cv. Banerjee's Giant) after ten days exposure to elevated CO<sub>2</sub> (600 ± 50 ppm)

TABLE 8: EFFECT OF ELEVATED CO<sub>2</sub> (600 ±50 ppm) ON FOLIAR STARCH CONTENT IN SPINACEA OLERACEA

Sample		Starch Content (%)
S. oleracea (cv. All Green)	Control	2.3
	Elevated CO <sub>2</sub>	4.0
S. oleracea (cv. Banerjee's Giant)	Control	1.2
	Elevated CO <sub>2</sub>	2.3

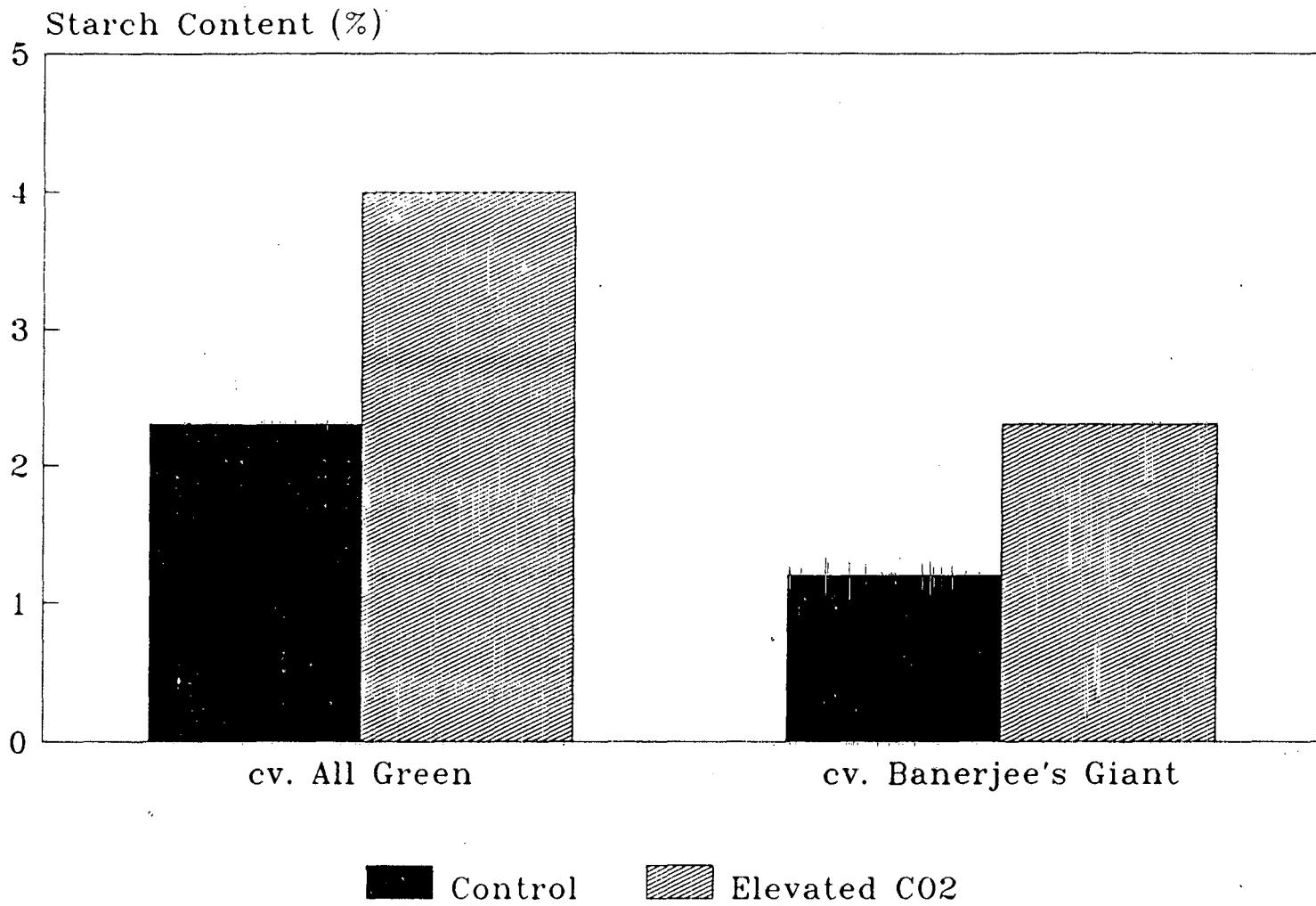


Fig 7 Percentage increase in foliar starch content after ten days exposure to elevated CO<sub>2</sub> (600 ± 50 ppm)

## Discussion

The leaf area in Spinacea oleracea plants exposed to elevated CO<sub>2</sub> (600 ± 50 ppm) for ten days continuously exhibit a marked increase over control plants. The percentage increase in leaf area in S. oleracea cv. All Green under elevated CO<sub>2</sub> was 94.2%, and in S. oleracea cv. Banerjee's Giant it was 40.14%, over control. Under elevated CO<sub>2</sub> enhanced photosynthetic rates promoted the growth of additional leaf area. Individual plants vary with respect to % increase in leaf area under elevated CO<sub>2</sub>. Increase in leaf area has also been recorded for other C<sub>3</sub> species as G. max (Rogers et. al., 1986), G. hirsutum (Delucia et.al, 1985). P. vulgaris, plants (Porter and Grodzinski, 1984) showed a 28% increase in leaf area under CO<sub>2</sub> enrichment (1200 ppm) compared to control over a 14-day exposure period.

Total chlorophyll content declined under elevated CO<sub>2</sub>, 29.75% reduction in S. oleracea cv. All Green, 6.97% in S. oleracea cv. Banerjee's Giant, over control. Total chlorophyll has been reported to decline under CO<sub>2</sub> enrichment in other plants too. A, 42% reduction has been reported in G. hirsutum (Chang, 1975), 30% in T. subterraneum (Cave et al., 1981), 61% in G. hirsutum (Delucia et al., 1985), 14% in C. album (Sage et al., 1989) and 34% in B. oleracea (Sage et al., 1989). Reduction in

chlorophyll content at elevated CO<sub>2</sub> is attributed to chloroplast degeneration as a result of excess starch accumulation (Madsen, 1968). Chl a:b ratio increased under elevated CO<sub>2</sub> in S. oleracea cv. All Green from 1.68 to 2.11 and from 2.3 to 3.05 in S. oleracea cv. Banerjee's Giant. This result is in contrast to values reported in literature where chl a:b ratio has been found to decrease under elevated CO<sub>2</sub> (Cave et al., 1981; Delucia et al., 1985). The reason for the observed increase in chl a:b ratio could be, a greater % reduction in chl (b) compared to chl (a), under CO<sub>2</sub> enrichment. Carotene was also found to decrease under CO<sub>2</sub> enrichment. The information about response of carotene under enhanced CO<sub>2</sub> is practically non existent.

An increase in root and shoot biomass was observed in plants exposed to elevated CO<sub>2</sub>. Percentage increase in root biomass was more than shoot biomass in both cultivars, with S. oleracea cv. All Green exhibiting greater % increase than S. oleracea cv. Banerjee's Giant. Similar results have been obtained in G. max (Cure et al., 1987) where, by day 22 of exposure to 700 ppm CO<sub>2</sub> biomass of roots increased by 88% over control as compared to 60% in leaves and 73% in stems. But contrasting results have been obtained in G. hirsutum (Delucia et al., 1985) where biomass partitioning was preferential to leaf sink followed by stems and least to roots. Root/Shoot ratio was found to increase under elevated

CO<sub>2</sub> from 0.161 to 0.348 (S. oleracea cv. All Green) and from 0.096 to 0.160 (S. oleracea cv. Banerjee's Giant). The results indicate that there is a preferential partitioning of photosynthates to root system. Ambient CO<sub>2</sub> thus, seems to have a definite control over photosynthate partitioning. Generalizations are not possible since the response seems to be species specific.

Foliar starch content increased in both cultivars with cv. All Green exhibiting a greater % starch content (4%) under elevated CO<sub>2</sub> (4%) compared to control (2.3%) than in cv. Banerjee's Giant (2.3 & 1.2%). In G. max, Finn and Brun (1982), reported a 46% increase in leaf starch content. In T. subterraneum (Cave et al., 1981), the percentage increase in starch content under elevated CO<sub>2</sub> was 135% compared to control (46.7%). The reason for lower leaf starch content in S. oleracea under elevated CO<sub>2</sub> could be, that additional photosynthate is present as sugars rather than starch.

The results clearly indicate that S. oleracea being a C<sub>3</sub> plant responds positively to elevated CO<sub>2</sub> as expected. But distinct intervarietal difference in response of S. oleracea to elevated CO<sub>2</sub> are evident (Plate 3,4). S. oleracea cv. All Green is more responsive to elevated CO<sub>2</sub> levels than S. oleracea cv. Banerjee's Giant, as the



Plate 3 *S. oleracea* cv. All Green plants after ten days exposure to A.  
Ambient CO<sub>2</sub> B. Elevated CO<sub>2</sub> (600 ± 50 ppm)





Plate 4 *S. oleracea* cv. Banerjee's Giant plants after ten days exposure to A. Ambient CO<sub>2</sub> B. Elevated CO<sub>2</sub> (600 ± 50 ppm)

former showed a greater % increase in leaf area, biomass and starch content under elevated CO<sub>2</sub>. Percentage reduction in chlorophyll and carotene content was also greater in S. oleracea cv. All Green under elevated CO<sub>2</sub>.

Studies using open top chamber have not been done in India so far. In absence of any previous experience, chamber construction and standardization took a major part of the time available. In addition to this CO<sub>2</sub> was monitored rigorously every three hr. Due to these demanding conditions and limited time available the exposure period could not be extended beyond ten days and replicates also could not be obtained. As a result the data could not be subjected to statistical analysis. In spite of these limitations definite trends in response of S. oleracea to enhanced CO<sub>2</sub> were quite discernible. These results can only be treated as indicative rather than being conclusive in characterisation of response of S. oleracea to elevated CO<sub>2</sub>. In future, long term studies on response of local plants to enhanced CO<sub>2</sub> levels using standard open top chambers are important. Such studies are required to obtain basic information for making a more realistic evaluation of impact of elevated CO<sub>2</sub> on plants.

## ABSTRACT

Eighteen - day old Spinacea oleracea cv. All green and S. oleracea cv. Banerjee's Giant plants were exposed to ambient ( $350 \pm 25$  ppm) and elevated ( $600 \pm 50$  ppm)  $CO_2$  in open - top chambers over a ten day period continuously. Leaf area increased by 94.2% (S. oleracea cv. All Green) and 40.14% (S. oleracea cv. Banerjee's Giant), under elevated  $CO_2$  over ambient  $CO_2$  plants. Root and shoot biomass increased in both cultivars under elevated  $CO_2$ , 296 and 83.5% (S. oleracea cv. All Green), 188.8 and 73.5% (S. oleracea cv. Banerjee's Giant) respectively. Root : Shoot ratio increased from 0.161 to 0.348 (S. oleracea cv. All Green) and from 0.096 to 0.160 (S. oleracea cv. Banerjee's Giant) under elevated  $CO_2$ . Foliar starch content increased slightly. Total chlorophyll content decreased by 29.75% (S. oleracea cv. All Green) and 6.97% (S. oleracea cv. Banerjee's Giant) compared to control. Chl a:b ratio increased under  $CO_2$  enrichment. Enhanced photosynthetic rates leading to greater dry matter production under elevated  $CO_2$  seem to be responsible for the observed increases in leaf area, biomass and starch. Decline in chlorophyll content could be due to disruption of chloroplasts by excess starch accumulation (Madsen, 1968). Out of the two cultivars of S. oleracea, studied, S. oleracea cv. All Green seems to be more responsive to

elevated CO<sub>2</sub> levels than S. oleracea cv. Banerjee's Giant. Overall positive responsive of S. oleracea may well enhance the market value of this leafy vegetable under enriched CO<sub>2</sub> atmospheres. The present study was of preliminary, short-term investigatory nature. Long-term, indepth study is needed to substantiate the results obtained above.

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